

MOLECULAR BASIS OF ETHNIC DIFFERENCES IN DRUG DISPOSITION AND RESPONSE

Hong-Guang Xie, Richard B Kim, Alastair JJ Wood,
and C Michael Stein

*Division of Clinical Pharmacology, Departments of Medicine and Pharmacology,
Vanderbilt University School of Medicine, Nashville, Tennessee 37232-6602;
e-mail: hong-guang.xie@mcmail.vanderbilt.edu, richard.kim@mcmail.vanderbilt.edu,
alastair.wood@mcmail.vanderbilt.edu, michael.stein@mcmail.vanderbilt.edu*

Key Words ethnic variation, drug metabolizing enzymes, drug transporters, drug receptors, genetic polymorphisms, population pharmacogenetics

■ **Abstract** Ethnicity is an important demographic variable contributing to interindividual variability in drug metabolism and response. In this rapidly expanding research area many genetic factors that account for the effects of ethnicity on pharmacokinetics, pharmacodynamics, and drug safety have been identified. This review focuses on recent developments that have improved understanding of the molecular mechanisms responsible for such interethnic differences. Genetic variations that may provide a molecular basis for ethnic differences in drug metabolizing enzymes (CYP 2C9, 2C19, 2D6, and 3A4), drug transporter (P-glycoprotein), drug receptors (adrenoceptors), and other functionally important proteins (eNOS and G proteins) are discussed. A better understanding of the molecular basis underlying ethnic differences in drug metabolism, transport, and response will contribute to improved individualization of drug therapy.

INTRODUCTION

The term ethnicity is a multidimensional classification that encompasses shared origins, social background, culture, and environment (1, 2). Ethnicity is an important determinant of drug metabolism and response and therefore contributes to interindividual variability. The definition of ethnicity, encompassing both genetic and environmental factors, is different from that of race (2). It is generally recognized that the effects of ethnicity on drug metabolism and response are determined by both genetic and environmental factors to a varying extent, depending on the ethnic groups and probe drugs studied (3–8a). Increased research in population pharmacogenetics (3–13) has led to the coining of terms “pharmacanthropology” (14) and “ethnopharmacology” (11).

DRUG METABOLIZING ENZYMES

CYP2C Subfamily

In human liver microsomes the cytochrome P-450 (CYP) 2C subfamily is second in quantity only to the P-450 3A subfamily (15). Its four known members are CYP 2C8, 2C9, 2C18, and 2C19 (16, 17). Of these, CYP 2C9 and 2C19 are the predominant *CYP2C* gene products. Comparison of the amino-terminal amino acid sequence of CYP2C9 shows that it differs from CYP2C19 by only two residues (16). Genetic variations in both CYP2C9 and 2C19 have clinical significance.

CYP2C9 CYP2C9 contributes ~20% of total hepatic P-450 content (18) and is one of the important drug metabolizing enzymes in humans (19). It has many clinically relevant substrates, including the oral anticoagulants warfarin and acenocoumarol, the oral hypoglycemics tolbutamide and glipizide, the anticonvulsant phenytoin, the loop diuretic furosemide, the angiotensin II-receptor antagonist losartan, the HMG-CoA inhibitor fluvastatin, and a number of nonsteroidal anti-inflammatory drugs (NSAIDs) (16, 19).

An early study of tolbutamide metabolism suggested that ~30% of subjects were poor metabolizers (PMs) (20). However, many subsequent studies, including a survey of tolbutamide oxidation capacity in 106 unrelated Australian participants that failed to find a single PM (21), indicated that the frequency of the CYP2C9 PM phenotype is low (22–29).

The gene encoding CYP2C9 protein is localized on chromosome 10 (30), has 9 exons, and is ~55 kb in size (GenBank accession numbers: L16877–L16883) (31). *CYP2C9* cDNA encodes a protein of 490 amino acids. There appear to be at least two naturally occurring variants: the wild-type Arg¹⁴⁴Ile³⁵⁹ (designated *CYP2C9**1), Cys¹⁴⁴Ile³⁵⁹ (designated *CYP2C9**2), and Arg¹⁴⁴Leu³⁵⁹ (designated *CYP2C9**3). The *CYP2C9**2 allelic variant has an exchange of C⁴³⁰ → T in exon 3, and *CYP2C9**3 has an exchange of A¹⁰⁷⁵ → C in exon 7. In vitro studies have shown that the allelic variants of *CYP2C9* differ in their affinity (K_m) and/or intrinsic clearance (V_{max}/K_m) for different substrates of CYP2C9 (21, 24, 32–35). For example, the *CYP2C9**2 variant was found to be associated with impaired 6- and 7-hydroxylation of *S*-warfarin (21, 32). However, other studies found that this variant resulted in a small or negligible decrease in the V_{max} for tolbutamide (21, 24, 32, 36), did not alter CYP2C9-mediated methyl hydroxylation of furosemide (34), and had little effect on *S*-warfarin hydroxylation (36, 37). In clinical studies individuals heterozygous for *CYP2C9**1/*2 required a 20% lower mean maintenance dose of warfarin to maintain therapeutic anticoagulation than wild-type homozygotes (38). Individuals homozygous for *CYP2C9**2 had 58.2% higher mean phenytoin trough levels than *CYP2C9**1 homozygotes (26).

The *CYP2C9**3 variant has been found to have a pronounced reduction in catalytic activity across all CYP2C9 substrates. In vitro evidence revealed that the product of the *CYP2C9**3 variant had a significantly lower maximum catalytic rate

(with lower V_{\max} value) and/or lower affinity (with higher K_m value) for *S*-warfarin, tolbutamide, and phenytoin than the wild-type form (18, 21, 24, 33, 36, 39). Individuals homozygous for *CYP2C*3* have been identified to be PMs of *S*-warfarin (25), phenytoin (23, 26, 40), glipizide (40), tolbutamide (23, 24, 41), and losartan (23, 27). Also, in individuals heterozygous for *CYP2C9*3* decreased metabolic clearance of phenytoin (26, 42, 43), *S*-warfarin (25, 28, 29, 38, 44, 45), and acenocoumarol (45a) was reported. Thus, the *CYP2C9*3* variant is a major genetic determinant of variability in the disposition of *CYP2C9* substrates.

The relationship between ethnicity and *CYP2C9* expression or activity is less clear. Using an immunochemical approach to quantify P-450 content, Shimada et al found that P-450 2C (principally *CYP2C9*) was not different in Caucasian and Japanese liver microsomes ($n = 30$ each) (15). A limited number of in vivo studies suggested that African Americans had slower hepatic metabolism of oral phenytoin than Caucasian Americans (46). Caucasians (American or German) had a higher mean K_m value (5.7–6.8 $\mu\text{g/ml}$; here estimated from population pharmacokinetic data) for phenytoin than did East Asians (Chinese or Japanese) (2.2–3.2 $\mu\text{g/ml}$) (47), but there are few studies comparing the disposition of *CYP2C9* substrate drugs in different ethnic groups.

More recently, analysis of *CYP2C9* polymorphisms has been performed across various ethnic groups. The allelic frequencies of both *CYP2C9* variants (*2 and *3) in different ethnic groups are summarized in Table 1. The *CYP2C9*2* variant is present only in black (~3%) and white (~10%) populations and is absent or

TABLE 1 Ethnic distribution of the *CYP2C9* allelic variants^a

Ethnicity	Cys ¹⁴⁴ (*2)		Leu ³⁵⁹ (*3)		References
	n	%	n	%	
Asians					
Chinese	466	0.0	426	2.1	24, 48
Japanese	1394	0.0	1394	1.9	18, 42, 44, 49, 50 ^b
Korean	1148	0.0	1148	1.1	47a
Total	3008	0.0	2968	1.6	
Blacks					
American	1098	2.9	500	0.8	24, 51 ^b
Caucasians					
American	370	10.0	1512	7.9	18, 24, 51 ^b
British	588	14.1	400	9.5	28, 38, 52
German	988	11.3	734	7.8	53, 54
Swedish	860	10.7	860	7.4	55
Turkish	998	10.6	998	10.0	26
Total	3804	11.3	4504	8.4	

^an, combined number of the total alleles tested; %, percent of the allelic variants.

^bUI Schwarz, EF Choo, GK Dresser, CM Stein, AJJ Wood, DM Roden, GR Wilkinson, RB Kim. 2000, unpublished data.

extremely rare in East Asians (18, 24, 26, 28, 38, 42, 44, 47a, 48–55), suggesting that any effects of this variant on CYP2C9-mediated drug metabolism are likely to be negligible in East Asians. White subjects from many parts of the world have a significantly higher frequency of both *CYP2C9**2 and *3 (~10%) than Asian or black subjects. Asian individuals homozygous for *CYP2C9**3 were rare, and less than ~2% of the Asians were heterozygous for this allelic variant. Because of both the greater frequency of the *CYP2C9**3 allele in white (~8%) than East Asian (~2%) or black (~1%) populations and the effects of gene dosage on CYP2C9 activity (26, 28, 45), we would anticipate that concentrations of CYP2C9 substrate drugs would, on average, be higher in whites, assuming expressed levels of the protein are comparable.

Ethnicity affects the average warfarin dose required to maintain therapeutic anticoagulation (56). However, in contrast to what would be predicted from ethnic differences in the frequency of the *CYP2C9**3 allele, white patients require higher warfarin doses than Asians to attain a comparable anticoagulant effect. Chinese patients required a ~50% lower average maintenance dose of warfarin (3.3 mg/day) than white patients (6.1 mg/day) to obtain comparable anticoagulation [international normalized ratio (INR): 2.0–2.5] (57). The average maintenance dose of warfarin for Japanese patients with heart disease (3.3 mg/day) is also much lower than that for American patients (4.9 mg/day) (44). These data imply that known CYP2C9 polymorphisms account for only part of the ethnic differences in sensitivity to warfarin. Similarly, because of a higher frequency of *CYP2C9**3, we would anticipate that whites would have a higher incidence of hypoglycemia induced by tolbutamide or glipizide, and a higher risk of warfarin-induced bleeding complications (28, 28a) than blacks or Asians taking similar doses; there is, however, little information regarding the effect of ethnicity on response to these drugs.

Recently, an Asp(GAC)³⁶⁰Glu (GAG) polymorphism was identified in 5 of 110 African Americans with an allele frequency of 2.3% (58). Interestingly, another new *CYP2C9* variant ATT (Ile³⁵⁹) → ACT (Thr³⁵⁹) was identified in 32 Japanese patients with epilepsy but not in 100 healthy Japanese subjects (59). Both polymorphic amino acid residues are close to the *CYP2C9**3 variant and lie within the putative substrate recognition site 5 (SRS 5) in the CYP2 family (60), but their functional significance is as yet unknown.

CYP2C19 Impaired 4'-hydroxylation of *S*-mephenytoin in humans is a good example of how polymorphisms can affect drug metabolism and alter clinical response (61–66). Most of the population in any ethnic group can be phenotyped as extensive metabolizers (EMs) based on their ability to oxidize *S*-mephenytoin or other CYP2C19 substrates. However, because of a metabolic defect resulting from genetic variations in *CYP2C19*, some individuals cluster outside of the normal distribution and can be classified as PMs. CYP2C19 is a clinically important enzyme that catalyzes the metabolism of several frequently prescribed drugs (16, 67) such as diazepam, some barbiturates, tricyclic antidepressants, proguanil, and omeprazole and its structural analogs.

CYP2C19 is a protein of 490 amino acids encoded by the *CYP2C19* gene, which has 9 exons (GenBank accession numbers: L31506, L31507, L32982, and L32983) (68) and is mapped to chromosome 10q²⁴ (69). CYP2C19 activity exhibits marked genetic polymorphism, and at least nine allelic variants have been identified (for the nomenclature, go to: <http://www.imm.ki.se/CYPalleles/cyp2c19.htm>). The two common alleles that result in a nonfunctional enzyme are null alleles. The first allelic variant, *CYP2C19**2 (previously designated *m1*), is a G⁶⁸¹ → A point mutation in exon 5 that introduces an aberrant splice site resulting in an alteration of the reading frame of the mRNA and a truncated nonfunctional protein (70). A second common defective allele, *CYP2C19**3 (previously termed *m2*), a G⁶³⁶ → A single base transition in exon 4, produces a premature stop codon (71, 72). These two allelic variants account for almost all PMs in East Asians and blacks (73–75), but in whites ~10% of the PM alleles have not yet been identified (73).

Several studies indicate that there are pronounced ethnic variations in the frequency of *CYP2C19* genotypes and phenotypes among different populations (reviewed in 12, 73–77). Recently, we have summarized global data on allelic, genotypic, and phenotypic frequencies of *CYP2C19* in white populations of European descent, black populations of African descent, and Chinese populations of Asian origin (73–75). As shown in Table 2, the frequency of *CYP2C19**2 in the Chinese population (30%) is twice that in black (17%) or white (15%) populations, and *CYP2C19**3 occurs in approximately 5% of Chinese but less than 1% of blacks or whites.

In addition to ethnic differences in the frequency of CYP2C19 polymorphisms, enzyme activity also varies. Zhang et al found no significant difference in diazepam clearance in Chinese EMs and PMs of *S*-mephenytoin (78). Both EM and PM phenotypes had a diazepam clearance that was comparable to the range observed in white PMs and was half that of white EMs (78–80). These data imply that many Chinese EMs are heterozygotes (80, 81). As shown in Table 2, within the EM subgroups, the proportion of heterozygotes in Chinese is approximately 50%, twice that in whites. The low clearance of diazepam in Chinese EMs (78) and anecdotal evidence that “many Hong Kong physicians routinely prescribe smaller diazepam doses for Chinese than for Caucasians” (82) could thus be explained by the higher

TABLE 2 Allelic, genotypic, and phenotypic *CYP2C19* frequencies in different ethnic groups (73–75)

	Phenotype		Genotype			Allele			
	n	PM (%)	n	PM (%)	wt/m (%) ^a	n	*1 (%)	*2 (%)	*3 (%)
Blacks	922	3.9	966	3.7	29.0	1932	82.3	17.3	0.4
Chinese	1555	13.6	573	13.8	49.8	1146	64.7	30.0	5.1
Caucasians	3990	2.8	1356	2.1	26.0	2712	85.3	14.7	0.04

^awt/m, the proportion of heterozygotes in the EM subgroups.

frequency of CYP2C19 PMs and heterozygous EMs in the Chinese population. Similarly, the higher proportion of *CYP2C19**2 and *3 heterozygotes in Chinese EMs may be the molecular explanation for the observation that Caucasian EMs were more efficient at metabolizing omeprazole than Chinese EMs (62, 83–86) or Koreans (86). CYP2C19 also appears to be the major enzyme that activates the antimalarial drug proguanil to produce its therapeutically active metabolite cycloguanil (87). However, failure of proguanil's malaria chemoprophylaxis was not more likely in populations with a higher frequency of CYP2C19 PMs and *CYP2C19**2 and *3 heterozygotes, such as in Melanesians living in the Vanuatu islands, where malaria is endemic (88), suggesting that metabolic pathways other than CYP2C19 may be important or that the parent compound proguanil has significant intrinsic efficacy against malaria, independent of the active metabolite, cycloguanil.

CYP2D6

CYP2D6 is of clinical relevance because the gene encoding this enzyme is highly polymorphic (for more information, go to <http://www.imm.ki.se/CYPalleles/cyp2d6.htm>). At least 70 *CYP2D6* alleles are responsible for the ~200-fold variability in the metabolism of 100 or more drugs (89). Among the common allelic variants, the functional alleles include *CYP2D6**1 (wild-type), *2, *9, *10, and *17, whereas almost all the remainder are nonfunctional. Several mechanisms are responsible for genetic variability in *CYP2D6*. These include whole gene deletion (e.g. *CYP2D6**5, a 12.1-kb deletion that includes the entire *CYP2D6* gene), gene duplication or multiplication [e.g. *CYP2D6**1 \times n; *2 \times n (n = 1, ..., 13); *4 \times 2, and *35 \times 2], single nucleotide polymorphisms alone or in combination, deletion or insertion of single or multiple base(s), gene conversion (e.g. *CYP2D6**36), and repeats (e.g. *CYP2D6**30). Most of the known variant alleles are inactive and produce the PM phenotype, which appears to be rare (~1%) in most Asian populations, more common in whites (5%–10%) (12, 13, 90), and varies in black populations of African descent (0%–19%) (8, 91). Genotype-phenotype relationship studies have demonstrated that determination of the seven most common allelic variants, *2, *3, *4, *5, *6, *9, and *10, resulted in a correct phenotype assignment for nearly 100% of all patients of European descent (92, 93). Based on the known *CYP2D6* genotype-phenotype relationships, individuals can be phenotyped into four potential subgroups: ultrarapid metabolizers, extensive metabolizers (EMs), intermediate metabolizers, and poor metabolizers (PMs). Theoretically, ultrarapid metabolizers carry at least three active alleles, and the molecular bases for this phenotype are functional alleles with more than one gene copy; EMs are defined by the presence of the two functional *CYP2D6* alleles (*1, *2, *9, *10, and *17); intermediate metabolizers carry only one active allele (frequently *2, *9, or *10); and PMs lack functional *CYP2D6* alleles. Compared with the wild-type allele *1, active alleles *2, *9, *10, and *17 are functional but have a moderately decreased CYP2D6 enzyme activity (93–102).

Significant ethnic differences have been demonstrated in: (a) the population frequency of the PM phenotype; (b) the distribution patterns of the metabolic ratio (MR) within individuals with the EM phenotype; (c) in vitro and in vivo metabolism of CYP2D6 substrates; (d) phenotype-genotype correlations; (e) the effects of gene-dosage on the metabolism and response to drugs that are substrates for CYP2D6; and (f) the population frequency of allelic variants.

In an in vitro study, Caucasian liver microsome preparations tended to have higher CYP2D6 content and higher bufuralol (a substrate of CYP2D6) 1'-hydroxylase activity than microsomes of Japanese origin ($n = 30$ each) (15). The frequency distribution of the debrisoquine MRs in Chinese EMs was shifted to the right (higher values) compared with Swedish EMs. Most Chinese EMs (~66%) had a MR >1, whereas a smaller proportion of Swedish EMs (~20%) had a MR >1 (103), and the hydroxylation of debrisoquine was slower in Chinese EMs than white EMs (104). This ethnic difference would be expected to also occur with other CYP2D6 substrates. For example, the mean clearance of desipramine, a substrate of CYP2D6, was considerably lower in Chinese EMs than in white EMs (105). Also, the production of morphine from CYP2D6-catalyzed *O*-demethylation of codeine was lower in Chinese EMs than Caucasian EMs (106–108). Recently, we demonstrated that codeine's apparent clearance and partial metabolic clearance via *O*-demethylation are significantly higher in white American EMs than in Chinese EMs, and that when codeine was co-administrated with quinidine (a potent inhibitor of CYP2D6) the inhibition of the production of codeine *O*-demethylated metabolite, morphine, is markedly greater in whites than in Chinese. The respiratory depressant effect caused by morphine after administration of codeine is significantly greater in Caucasians than in Chinese, before and after quinidine (108). These data suggest that although East Asians rarely have the CYP2D6 PM phenotype, within the EM population they have lower levels of CYP2D6 activity than Caucasian EMs.

Among different East Asian ethnic groups significant variations in CYP2D6 activity exist. Ishizaki and colleagues used metoprolol as a probe for CYP2D6 phenotyping and observed that the distribution of metoprolol MRs in Chinese EMs was also shifted to the right compared with Japanese or Korean EMs (109–110). The distribution of the metoprolol MRs in Korean EMs was similar to that in Japanese (110), whereas the proportion of EMs with MRs >1 in Chinese (66%) (103) was twice that than in Koreans (~33%) (111). Consistent with the findings by both Horai et al (109) and Sohn et al (110), Chinese have a significantly lower ability to *O*-demethylate codeine than Japanese and Koreans (112). The allele *CYP2D6**10 is a Pro³⁴ → Ser substitution in the proline-rich region near the NH₂-terminal that results in impaired folding capacity of the enzyme so that although it is functional, it has decreased activity (96). As shown in Table 3, *CYP2D6**10 is more frequent in East Asians such as Chinese (51–70%), Japanese (~40%), and Koreans (51%) than in either whites (1–7%) or blacks (1–9%) (92–94, 96, 100, 111, 112a, 113–118, 121, 123, 124, 126, 131, 134). Thus, the more common occurrence of allele *10 is likely to contribute to the decreased CYP2D6 activity in East Asians. The

TABLE 3 Ethnic distribution of the major *CYP2D6* alleles^a

	n	*1	*2	*3	*4	*5	*6	*9	*10	*17	*M × N	References
Asians												
Chinese	248			0.0	0.8	1.2			70.0		0.9	113
	226	26.9	13.4			5.7			50.7		1.3	96
	238	22.7	8.0		0.0	4.6			64.7			114
Japanese	196	42.3	9.2		0.5	6.1			40.8			100
	324	40.1	13.0	0.0	0.0	6.2			38.6	0.0		115
	412	43.0	12.3		0.2	4.5			38.1		1.0	112a
Korean	304	49.0							51.0		0.3	111
Blacks												
American	88	28.0	24.0	1.0	5.0	9.0			1.0	29.0	1.0	116
	492	83.0 ^b		0.6	7.3	6.9			5.2	26.0	2.4	117, 118
	482	90.4 ^b		0.2	9.3							119
	254	84.6 ^b		0.4	9.1	5.9						120
Ethiopian	244			0.0	1.2	3.3			8.6	9.0	16.0	121
Tanzanian	216	27.8	40.0	0.0	0.9	6.3	0.0		3.8	17.0	3.4	94
	392			0.0	4.0						2.5	122
Zimbabwean	160	85.6 ^b	9.9	0.0	2.5	3.8		0.0	5.6	34.0	0.9	123, 124
Whites												
American	416	37.0	33.7	1.0	17.5	3.8	1.0	2.9	1.9	0.2	1.1	93
	112				17.8	0.9						125
	928	76.0 ^b		1.2	18.1	2.9			4.0	0.0	2.3	117, 118
	928	78.3 ^b		1.0	20.7							119
	252	66.7 ^b		2.4	28.6	2.4						120
British	1332	33.4	32.9	1.8	18.9	7.3	1.4	2.6	1.4	0.1		126
Danish	650	76.8 ^b		2.0	20.6	0.6						127
	480	72.3 ^b		2.5	18.1	5.2						128
Estonian	302	76.2 ^b		2.3	21.5						0.7	129
Finnish	604	81.5 ^b		2.2	12.8	1.7		0.3			1.2	130
German	390	35.6	33.6	1.0	19.5	4.1	1.3	2.1	2.1	0.0	1.6	131
	1154	36.4	32.4	2.0	20.7	2.0	0.9	1.8	1.5		1.9	92
Russian	408	83.8 ^b		1.2	15.0							132
Swedish	496	75.0 ^b		2.0	23.0							133
	166	83.0 ^b		1.0	16.0							133
Turkish	808	37.1	35.3	0.0	11.3	1.5	0.7	0.6	6.1	1.1	5.6	134

^aData are presented as *n* and percent, where *n* = total number of the alleles tested. *M × N: duplication, amplification or multiplication of the alleles, referred to *1 × N or *2 × N, not including *4 × N because of lack of functional significance. Functional alleles are *1, *2, *9, *10, and *17; the remaining are nonfunctional.

^bOverestimated because the *CYP2D6**1 (wild-type) frequency was calculated from the number of polymorphic alleles detected. The empty cells mean that no data are available now.

significant difference in the frequencies of alleles *1 and *10 in Chinese and Japanese subjects might contribute to the ethnic differences in CYP2D6 activity levels between these two populations (96, 100, 112a, 113–115). The rarity of the inactive allele, *CYP2D6**4, in East Asians (100, 112a, 113–115) is thought to be associated with a low frequency of the PM phenotype (~1%). The active allele *CYP2D6**2 has frequently been found to be the variant associated with gene duplication or multiplication and thus significantly increased levels of CYP2D6 activity (134a). The lower frequency of the *2 allele could also contribute to the lower enzyme activity in East Asians (96, 100, 112a, 114, 115) compared with whites (92, 93, 126, 131, 134).

In black African populations the frequency of the CYP2D6 PM phenotype has varied in different studies (0–19%), depending on the ethnic group and probe drug studied (8, 91). In EMs a high proportion of individuals had a debrisoquine MR >1 in Zimbabwe (49–59%) (94, 123, 124) and Ethiopia (37%) (121). Marked variations in CYP2D6 activity are present among different black African ethnic groups, but overall, CYP2D6 activity is lower than in whites [e.g. ~21% of Swedish EMs had a debrisoquine MR >1 (135)]. Ethnic differences in the frequency of the *17 allele may explain these observations. The *CYP2D6**17 was found to have 20% of the wild-type CYP2D6 activity and altered affinity for CYP2D6 substrates such as bufuralol and codeine (99, 123). As shown in Table 3, although *CYP2D6**17 appears to be relatively African-specific (93, 94, 115–118, 121, 123, 124, 126, 131, 134), substantial variability in the *17 allele frequency occurs among different black African populations (9–34%) (94, 116–118, 121, 123, 124) and could partly explain the large variation in the frequency of the CYP2D6 PM phenotype (0–19%) reported in studies that used different probe substrates. In addition, variability in the frequency of the nonfunctional *4 allele (94, 116–124) may contribute to the varying frequency of the CYP2D6 PM phenotype in different black populations. Thus, the fact that CYP2D6 PMs are more common in Caucasians than in most black populations is likely to be due to the higher frequency of the nonfunctional *4 and *3 alleles in whites (92–94, 116–134), whereas the lower CYP2D6 activity in black EMs may be due to a significantly higher frequency of the *17 allele (see Table 3).

CYP3A4

The P-450 3A subfamily is the predominant P-450 isoform in human liver (~30% of total P-450 content) (15) and contains three known members (17), P-450 3A4, 3A5, and 3A7, that are expressed in several organs important in drug metabolism and disposition. CYP3A4 is abundantly present in human liver and small intestine (15, 136) and contributes to the metabolism of ~50% of commonly used drugs including nifedipine, cyclosporine, erythromycin, midazolam, alprazolam, and triazolam (136–141). It is important to note that interindividual variation in the levels of CYP3A4 expression is high, up to ~20-fold or more (15, 142–145), and may account for the wide range of interindividual variability in the disposition of drugs metabolized by this enzyme (145–149).

TABLE 4 Studies of CYP3A4 substrates in different ethnic groups

Substrate	Parameters	Results	References
Alprazolam	CL _o , CL _s	Lower in Asians than Caucasians	150
	CL _o , CL _s	Similar in American-born Asians and native Asians	150
Cerivastatin	AUC, C _{max}	Similar in blacks, Japanese, and Caucasians	151
Cyclosporine	PK	Similar in healthy African and Caucasian Americans	152
	AUC	Lower in African- than Caucasian-American patients	153, 154
Erythromycin	AUC, t _{1/2}	Similar in Koreans and Caucasians ^a	155
Midazolam	CL, CL/F, F	Similar in African- and Caucasian Americans	156
Nifedipine	AUC	Threefold higher in South Asians than Caucasians	157, 158
	CL _s	77% higher in Caucasians than Asian Indians	159
	AUC, t _{1/2}	Similar in Malaysians and Asian Indians	160
	AUC, t _{1/2}	1.4-fold higher in Koreans than Caucasians ^a	155
	C _{min,ss}	Similar in black West Indians and Caucasians	161
	AUC, t _{1/2}	Similar in Nigerians and South Asians	162
	AUC, t _{1/2}	81% higher in Nigerians than Caucasians ^a	162
Triazolam	CL _o , CL _m	Similar in Asian Indians and Caucasians	163

AUC, area under the plasma concentration–time curve with extrapolation to infinity; CL_o, oral clearance; CL_m, partial metabolic clearance; CL_s, systemic clearance; C_{max}, peak concentration; t_{1/2}, terminal half-life; F, bioavailability; PK, pharmacokinetics (parameters); C_{min,ss}, trough concentration at steady state.

^aNormalized for body weight.

Interethnic differences in CYP3A4-mediated drug metabolism have been studied in vitro and in different populations. Caucasian liver microsomes had higher nifedipine oxidase activity and significantly higher testosterone 6 β -hydroxylase activity than Japanese samples ($n = 30$ each). Hepatic P-450 3A (principally P-450 3A4) content correlated well with nifedipine oxidation ($r = 0.79$) and testosterone 6 β -hydroxylation ($r = 0.81$) activities. Also, CYP3A4-mediated metabolic activation of aflatoxin B1 and sterigmatocystin correlated well with microsomal P-450 3A content ($r = 0.78$ and 0.83 , respectively) and was significantly higher in Caucasian than in Japanese samples (15). As summarized in Table 4, the apparent oral clearance of alprazolam was found to be similar in native and American-born Asians, but to be significantly higher in Caucasians than in Asians (150). Similarly, the area under the plasma concentration–time curve of nifedipine was significantly higher in Asians than Caucasians (155, 157–159). The ability to metabolize oral nifedipine was similar in Asian Indians and Malaysians who resided in the same geographic area (160) and codeine *N*-demethylation, mediated by CYP3A4 (164), was more extensive in Caucasian than Chinese subjects (106). These data suggest that CYP3A4 activity may be higher in Caucasians than other populations. However, such an ethnic variation may be substrate-dependent, since erythromycin *N*-demethylation did not show a good correlation with the content of hepatic P-450 3A ($r = 0.28$) (15), and no difference was present between Asians and Caucasians when erythromycin (155), triazolam (163) or cerivastatin (151) were used as metabolic markers of CYP3A4 activity.

Conflicting results have been found when CYP3A4-catalyzed oxidation of nifedipine (161, 162) was compared in black and white subjects. The metabolism of cerivastatin (151) and midazolam (156) does not differ between blacks and whites. Based on published data (see Table 4), ethnic comparisons with different CYP3A4 substrates have yielded inconsistent results that are difficult to interpret and may reflect an interplay of genetic and environmental factors.

An early in vivo study of nifedipine oxidation showed an apparent bimodality of the area under the plasma concentration–time curve of nifedipine after a 20 mg oral dose in 53 healthy Dutch individuals (165). Subsequent studies, however, in a larger number of subjects, did not confirm this finding (137, 147, 148, 158, 166), suggesting that nifedipine's metabolism is unlikely to be governed by genetic variations that result in the presence or absence of enzyme activity, such as occurs with CYP2C19 and CYP2D6.

The *CYP3A4* gene was mapped on chromosome 7q^{22.1} (167–171) and found to be ~27 kb long, with 13 exons and 12 introns (172). The sequence of *CYP3A4* cDNA obtained from hepatic libraries has been extensively examined (GenBank accession numbers: M18907 for the *CYP3A4* cDNA, and D11131 for its 5'-flanking region) (168, 172–176). To date, at least four *CYP3A4* allelic variants have been identified (for more information on the nomenclature, go to <http://www.imm.ki.se/CYPalleles/cyp3a4.htm>). The first common *CYP3A4* allelic variant is an A to G transition in the 5' promotor region at position –290 (from the transcription initiation site) (177), altering the 10-bp nifedipine-specific response element localized at –287 to –296 of the 5' regulatory region (172). This allele (previously termed *CYP3A4-V* and now designated as *CYP3A4*1B*) was found to be more common in patients with prostate cancer of a more invasive clinical stage than patients with a low-level clinical stage (177), and to be over-represented in patients with leukemia (178). Recent studies indicate that this allelic variant results in a modest reduction in hepatic CYP3A4 activity (156) but does not significantly alter the metabolism of CYP3A4 substrate drugs (179–181), although there are marked differences in frequency among various ethnic populations (181a). The frequency of *CYP3A4*1B* was low in white and Hispanic subjects (3.6–11.0%) (177, 179, 181a, 182), absent in Chinese and Japanese subjects (179, 182, 183), and much higher in black subjects (53.0–69.0%) (179, 181a, 182, 183).

A second allelic variant of the *CYP3A4* gene was found in exon 7 (a Ser²²²Pro substitution) and designated *CYP3A4*2* (183). This allele was uncommon in 55 white subjects (2.7%), and was not observed in black or Chinese groups of similar size (183). Using a baculovirus-directed cDNA expression system, the intrinsic clearance (V_{\max}/K_m) for nifedipine oxidation was decreased approximately six- to ninefold with the variant enzyme compared with the wild-type enzyme, but was not significantly different for testosterone 6 β -hydroxylation (183). The *CYP3A4*2* allele may encode a variant enzyme with substrate-dependent altered kinetics, but because it is uncommon, this variant is not likely to contribute substantially to ethnic variations in CYP3A4 activity. Taken together, ethnic differences in the disposition of CYP3A4 substrate drugs are poorly characterized and inconsistent, and currently recognized molecular variations in the *CYP3A4* gene do not appear to

contribute substantially to interindividual variability in the disposition of CYP3A4 substrate drugs.

DRUG TRANSPORTER

P-Glycoprotein

There is increasing evidence that drug metabolism alone does not account for the observed interindividual variability in drug disposition or response (184), but that other processes, including drug transport, are important determinants of drug disposition. Although a number of drug transporters have been shown to play a key role in drug disposition (184–186; RB Kim & GR Wilkinson, submitted for publication), P-glycoprotein (P-gp), the *MDR1* gene product, is one of the best studied and characterized.

Although initial interest in P-gp focused on its role as a mediator of multidrug resistance (MDR) in tumor cells, recent studies have demonstrated its more general role in drug disposition (184–188). P-gp is expressed in many tissues other than tumor cells, particularly those associated with excretory function (189), such as the canalicular domain of hepatocytes and the (luminal) brush border membrane of both intestinal epithelial cells and renal proximal tubule cells. P-gp is an ATP-dependent drug efflux pump, actively transporting many structurally diverse compounds from the inside to the outside of cells against a concentration gradient. Its apical distribution in cells results in decreased drug absorption from the gut lumen and enhanced drug excretion into bile and urine. Moreover, expression of P-gp in the capillary endothelium of the blood-brain barrier prevents penetration of substrate drugs into the central nervous system. Accordingly, P-gp plays an important role in drug absorption, distribution, and excretion. Of importance for drug disposition is that P-gp and CYP3A4 are frequently co-expressed in the same cells and share a large number of substrates and modulators (139). The disposition of such drugs is thus affected by both transport and metabolism (184).

Variability in P-gp-mediated drug transport in the gastrointestinal tract alters the oral bioavailability (F) of P-gp substrates (7, 186, 190). Such effects are seen most easily when either hepatic or extrahepatic drug metabolism is negligible (188). Ethnic variation in P-gp activity has not been widely studied. Lindholm et al investigated the effects of demographic factors on the pharmacokinetics of cyclosporine, a drug that is a substrate for both P-gp and CYP3A4, in 187 kidney transplant recipients, and found that the oral bioavailability of cyclosporine was significantly lower in blacks ($n = 58$, mean = 30.9%) than whites ($n = 86$, mean = 39.6%) or Hispanics ($n = 40$, mean = 42.1%), with no ethnic variation in clearance and volume of distribution at steady state (V_{ss}) (153). Because a 10-fold variation in the levels of intestinal CYP3A4 had no clear effect on oral cyclosporine pharmacokinetics, Lown and colleagues postulated that intestinal P-gp transport activity was the major determinant of bioavailability and

C_{\max} of cyclosporine (190), so that patients with lower levels of intestinal P-gp had higher bioavailability and higher C_{\max} , and vice versa. Furthermore, the concentration/dose ratio of tacrolimus (also a substrate of CYP3A4 and P-gp) was correlated with the mRNA expression of MDR1 but not CYP3A4 (191). Like cyclosporine (153, 154, 192, 193), higher doses of tacrolimus were required in blacks than whites to attain similar plasma levels (194), suggesting a lower oral bioavailability of tacrolimus. Although there is no supporting evidence, one explanation for the lower bioavailability of cyclosporine and tacrolimus would be greater P-gp-mediated drug transport in blacks. We found no difference in the disposition of cyclosporine in healthy black and white men studied on a controlled diet (CM Stein, AJ Sadeque, JJ Murray, C Wandel, RB Kim, et al, manuscript submitted). Recently, we found that the oral clearance of fexofenadine (a P-gp substrate that is not metabolized) exhibited ~ 10 -fold interindividual variation and tended to be slightly higher in white women than in black women (195). However, additional studies with larger sample sizes will be required to define the relationship between ethnicity and P-gp activity. The interrelationship between CYP3A4 and P-gp, and the effects of environmental factors such as diet, make defining ethnic differences in P-gp-mediated drug disposition difficult.

The *MDR1* gene encoding P-gp is located on chromosome 7q²¹ (196), with 28 exons encoding a protein of 1280 amino acids. Significant information about the structure-function analysis of P-gp has recently been summarized (197). Some naturally occurring polymorphisms of the *MDR1* gene have been found to correlate with potential clinical effects (198), or with the levels of intestinal *MDR1* expression and uptake of orally administered digoxin (a substrate of P-gp) (188). We are currently examining the hypothesis that allelic variants of *MDR1* might be associated with interindividual or interethnic variations in the disposition of P-gp drug substrates. Using PCR-based single-stranded conformational polymorphism (SSCP) methods, we found that a number of single nucleotide polymorphisms exist in multiple exons and the 5'-flanking promotor region of the *MDR1* gene in Japanese, black, and white American populations, and that these point mutations are distributed differently among the ethnic groups (198a). Indeed, genetic variability in *MDR1* appears to be more frequent than previously thought. Future genotype-phenotype relationship studies may provide additional insights into the role of P-gp as a determinant of interindividual or interethnic variability in drug response.

DRUG RECEPTORS

α -Adrenergic Receptor

The α -adrenergic receptor (α -AR) family comprises two subfamilies (α_1 -AR and α_2 -AR). Three subtypes of each have been identified pharmacologically and through molecular cloning: α_{1A} (formerly α_{1C}), α_{1B} , α_{1D} (formerly $\alpha_{1A/D}$), α_{2A} , α_{2B} , and α_{2C} (199, 200). Evidence suggests that the human α_{1A} -AR predominates

in arteries (201), whereas all three α_1 -AR subtypes (in particular α_{1A} and α_{1B}) are expressed in veins (201) and peripheral blood lymphocytes (202). The major α_1 -AR subtypes mediating vasoconstriction and regulating peripheral vascular resistance are α_{1A} and α_{1B} .

Several studies have demonstrated that blacks have greater vascular reactivity in response to α -adrenergic stimuli than whites (203–208). The vascular responses to intrabrachial artery infusion of phenylephrine (an α_1 -AR agonist) and to cold stress (a stimulator of endogenous norepinephrine release) were compared in African-American and Caucasian normotensive men (207–208) and α_1 -AR-mediated vasoconstrictor reactivity was significantly increased in blacks. The response in the superficial dorsal hand vein to local infusion of phenylephrine was reported to be blunted in normotensive blacks (209). The different results may be due to the site of drug action (artery versus vein), because the responsiveness of different types of blood vessel differs quantitatively (210).

Known polymorphisms of the human α_{1B} -AR are rare and appear to not be associated with the interindividual variations in response to phenylephrine (211, 212). Although African Americans had a significantly lower frequency of the amino acid variant Cys⁴⁹² of the α_{1A} -AR than Caucasian Americans, this polymorphism was not associated with hypertension and its effects on sensitivity to phenylephrine are not known (213). At present, the mechanisms underlying the increased α -adrenergic vascular sensitivity in African Americans are unknown.

β -Adrenergic Receptor

Three different β -adrenergic receptor (β -AR) subtypes have been cloned and pharmacologically characterized: β_1 , β_2 , and β_3 (200, 214). The presence of a putative fourth β -AR subtype (β_4) has been proposed based on recent pharmacological studies in human and rat cardiac tissue (200). Although both β_1 -AR and β_2 -AR subtypes co-exist in the human cardiovascular system (200, 215), β_1 -ARs predominate. Many studies have revealed that β_1 -AR-mediated effects include exercise-induced increase in heart rate and systolic blood pressure, as well as renin release (for reviews, see 215–217), whereas β_2 -AR-mediated responses include a decrease in total peripheral resistance and diastolic blood pressure (215).

β_1 -Adrenergic Receptor (β_1 -AR) Ethnic differences in β -AR-mediated responses to drugs have been extensively investigated among Caucasians, East Asians, and blacks of African descent (4). Compared with Caucasian men, Chinese men had a greater sensitivity to the effects of propranolol, a nonselective β -AR antagonist, which produced a greater reduction in mean arterial blood pressure and exercise-induced tachycardia (218) and greater suppression of exercise-induced plasma renin release (a β_1 -AR-mediated effect) (219). By contrast, normotensive blacks had decreased sensitivity to isoproterenol (a nonselective β -AR agonist) compared with whites, before and after β -blockade with propranolol (220). Furthermore, clinical observations from many investigations have indicated that

black patients with hypertension respond less well to monotherapy with several β -AR blockers (221), including the nonselective β -blockers propranolol (222) and nadolol (223), and the β_1 -selective blocker atenolol (222–226). Decreased sensitivity to β -AR antagonists may be associated with the lower levels of plasma renin activity and higher proportion of low-renin hypertension found in blacks (4).

The human β_1 -AR is a protein of 477 amino acid residues encoded by an intronless gene (227, 228) localized on chromosome 10q^{24–26} (227). Recently, 18 single nucleotide polymorphisms have been identified in the human β_1 -AR gene, 17 of which are located in the *N*-terminal and *C*-terminal region of the coding exon, resulting in 7 amino acid substitutions (229). Two common allelic variants of the human β_1 -AR gene, A¹⁴⁵G (or Ser⁴⁹Gly) and C¹¹⁶⁵G (or Arg³⁸⁹Gly), were identified (228–234). The first variant Gly⁴⁹ was associated with a decreased mortality risk in patients with congestive heart failure (230) and was observed significantly more frequently in a group of patients with idiopathic dilated cardiomyopathy (229). However, no ethnic differences in the frequency of this allelic variant existed among African Americans, Caucasians, and Chinese (233). A second common variant Gly³⁸⁹ receptor was found to have a decreased receptor- G_s -protein interaction and reduced cyclic AMP production following exposure to agonist (232), suggesting that this variant receptor exhibits diminished response to a β_1 -AR agonist in vitro. In vivo studies to determine the functional significance of the Arg³⁸⁹Gly β_1 -AR polymorphism in humans are underway, and a preliminary population-based case-control study shows no association of this polymorphism with essential hypertension in African Americans or Caucasian Americans (234a). The frequency of the variant Gly³⁸⁹ receptor is significantly higher in African Americans (42%) than in Caucasian Americans (25%), Chinese (27%), or Hispanics (33%) (234b). The physiological significance of ethnic differences in the frequency of the Gly³⁸⁹ variant β_1 -AR, which is characterized as a loss-of-function polymorphism in vitro (232), is uncertain.

β_2 -Adrenergic Receptor (β_2 -AR) The human β_2 -AR is a protein of 413 amino acids that is encoded by an intronless gene mapped to chromosome 5q^{31–32} (235) and is distributed in the vascular smooth muscle cells of atria, ventricles, some arterioles (e.g. coronary and skeletal muscle vessels), and systemic veins (215). Selective β_2 -AR agonists produce vasodilation in humans. Thus, enhanced blood pressure responses to stress in blacks might be the result of blunted β_2 -AR-mediated vasodilation. In the human dorsal hand vein attenuated β_2 -AR-mediated vasodilation was observed in Asian Indians who resided in the United States compared with white Americans (236). We and others have compared forearm blood flow responses to isoproterenol in young black and white American normotensive men and found that responses were markedly blunted in blacks (220, 237–240). Endothelial release of NO has recently been found to contribute to the vasodilator effect of β_2 -AR stimulation (240–244). However, the vasodilator effect of isoproterenol was attenuated in normotensive black subjects both before and after *N*^G-monomethyl-L-arginine (an eNOS inhibitor) (240). Thus, the decreased vasodilator response

to isoproterenol in blacks is independent of the NO component of isoproterenol-induced vasorelaxation. The ethnic differences in β_2 -AR-mediated vasodilation raise the possibility that β_2 -AR polymorphisms may play a role.

Recently, two common naturally occurring allelic variants of the β_2 -AR, A⁴⁶G (or Arg¹⁶Gly) and C⁷⁹G (or Gln²⁷Glu), have been identified and their functional significance characterized (245). In contrast to the findings in vitro (246, 247), clinical studies showed that the Gly¹⁶ variant is resistant to isoproterenol-induced desensitization (248, 248a). The Gly¹⁶ variant, however, was associated with attenuated systemic vasodilation in response to intravenous infusion of a selective β_2 -AR agonist in normotensive Australian (249) and normotensive American white subjects (250). The Glu²⁷ β_2 -AR variant is resistant to agonist-stimulated β_2 -AR desensitization in vitro (246–248), and in vivo is associated with greater vasodilator responses to isoproterenol (248a, 251). There are ethnic differences in the distribution of the two common β_2 -AR polymorphisms (252, 253), with a significantly lower frequency of the variant Glu²⁷ allele, approximately 18%, in normotensive and hypertensive African Americans compared with Caucasian Americans (~35%). Whether the decreased frequency of the Glu²⁷ allele in blacks contributes significantly to their attenuated responses to β -AR agonists such as isoproterenol is currently under investigation.

OTHER FUNCTIONALLY IMPORTANT PROTEINS

Endothelial Nitric Oxide Synthase

Since the identification in 1987 of NO as a biological mediator, there has been an explosion of information about the physiological, pathophysiological, pharmacological, and therapeutic roles played by this molecule. NO is synthesized from L-arginine and molecular oxygen by the constitutive enzyme endothelial nitric oxide synthase (eNOS) that is expressed in the endothelium. NO diffuses from the endothelium into vascular smooth muscle cells, where it increases the levels of cGMP by stimulating soluble guanylate cyclase (GC), resulting in vascular relaxation (254). The release of NO by the endothelium contributes to basal vascular tone (255) and regulates blood flow and blood pressure (256). Recently, interethnic variations in NO-mediated responses to vasodilators such as acetylcholine, methacholine, bradykinin, and sodium nitroprusside, have been extensively assessed (239, 240, 257–259). Blacks were found to have markedly decreased NO-dependent vasodilator responses to acetylcholine (240, 258), methacholine (239), bradykinin (259), and sodium nitroprusside (an exogenous NO donor) (239, 240, 257), suggesting decreased cGMP-mediated vasorelaxation in blacks. In addition, blacks were found to have a reduced NO-dependent vasodilation during mental stress (257), and N^G-monomethyl-L-arginine significantly inhibited the stress-induced increase in the forearm blood flow in whites but not in blacks (257). Together, these data indicate that blacks have less endothelium-dependent and endothelium-independent NO-mediated vasodilation than whites.

The enzyme eNOS is encoded by the *NOS3* gene of 26 exons that is located on chromosome 7q³⁵⁻³⁶ (260). A common missense mutation was identified as a G⁸⁹⁴T single base exchange at the genomic position 1917 in exon 7 (or at position 894 in its cDNA sequence), producing an amino acid substitution (Glu²⁹⁸Asp). Preliminary in vivo observations suggest that acetylcholine-mediated, endothelium-dependent vasodilator responses are attenuated in healthy white Americans homozygous for the Asp²⁹⁸ variant compared with the wild-type homozygotes (261). The vasoconstrictor response to phenylephrine was also significantly higher in the variant 894T carriers (TT and GT) of French origin than in the GG carriers (262). These data suggest that the Asp²⁹⁸ (or 894T) variant of eNOS might be functionally important, resulting in decreased vasodilation. African Americans have a lower frequency of the variant Asp²⁹⁸ allele than Caucasian Americans (14.3% versus 35.3%) (263). Thus, this polymorphism would not explain the reduced vascular response to NO in African Americans.

Pertussis Toxin–Sensitive G_i-Type Protein

GTP binding proteins (G proteins) comprise a superfamily of ubiquitous signal-transducing proteins that participate in many intracellular signaling cascades and mediate the functional responses to numerous agonists. G proteins are heterotrimers with α , β , and γ subunits. A frequent genetic polymorphism (C⁸²⁵T) has recently been identified by Siffert et al (264) in exon 10 of the *GNB3* gene (chromosome 12p¹³) (265) encoding the β_3 subunit of pertussis toxin–sensitive G_i-type protein. This single nucleotide polymorphism is related to alternative splicing of exon 9, resulting in the loss of 41 amino acids, which results in increased sensitivity to agonists that stimulate intracellular signaling through the pertussis toxin–sensitive G protein (264). Because of the large number of receptors, including adrenoceptors that function through G protein interactions, functional polymorphisms might have important pathophysiological consequences.

Case-control studies suggest that the allelic 825T variant is associated with increased blood pressure in German (264, 266, 267) and Australian Caucasians (268) and black Caribbeans of West African descent (269), but not in Japanese (270–272), aboriginal Oji-Cree Canadians (273), French individuals (274), and African Americans (275), suggesting potential ethnic differences in the nature of genetic susceptibility loci. Furthermore, this variant was associated with left-ventricular hypertrophy in a Spanish (276) but not a German (277) population with hypertension. The 825T polymorphism was also associated with lower renin levels (266) and obesity in some studies (278–282a) but not in others (268).

α_2 -ARs are coupled to pertussis toxin–sensitive G_i protein and mediate vasoconstriction (283, 284). α_2 -AR-mediated coronary blood flow was significantly decreased in subjects with the *GNB3* 825T allele (285), but dorsal hand vein constrictor responses were not different (286). Individuals of African descent have a higher frequency of the 825T allele than Caucasians (79% versus 33%) (269, 278),

and the 825T allele was associated with hypertension in black Caribbeans of West-African descent (269) but not in African Americans (275).

SUMMARY AND FUTURE DIRECTIONS

Human P-450 enzymes associated with drug metabolism belong mainly to the CYP families CYP1, CYP2, and CYP3. The major forms (percent of total P-450 content) include CYP 3A (~30%), 2C (~20%), 1A2 (~13%), 2E1 (~7%), 2A6 (~4%), and 2D6 (~2%) (15). Approximately 40% of human P-450-mediated drug metabolism is catalyzed by polymorphic enzymes (134), and ~50% of commonly used drugs are metabolized by CYP3A4 (140). The greatest variability in the levels of enzyme activity is found with CYP 2D6 and 2C enzymes because of frequently occurring functionally significant polymorphisms (136). In this review, the relationship between such polymorphisms and ethnicity has been discussed. Other phase I and phase II drug metabolizing enzymes have been extensively reviewed elsewhere (11, 67, 287–290).

In addition, drug transporters (e.g. P-gp) function as drug efflux pumps in intestine, liver, and kidney and play an important role in drug absorption, distribution, and excretion. P-gp and CYP3A4 are commonly co-expressed in the same tissues and share substrate specificity. Thus, for many drugs both metabolism and transport are important determinants of disposition. Conclusive evidence for ethnic variations in the P-gp transporter activity is not available but is an intriguing possibility.

Ethnic differences in β -AR-mediated responses exist. Sensitivity to propranolol is greater in Chinese, and β_2 -AR-mediated vasodilation is attenuated in African Americans compared with Caucasians. There are ethnic differences in the distribution of functionally significant polymorphisms, but the molecular basis for ethnic differences in vascular response is undefined.

NO plays an important role in the regulation of basal vascular tone and vasodilation. Evidence suggests that African Americans have attenuated NO-mediated (both endothelium-dependent and -independent) vasodilation compared with white Americans. However, the genetic or environmental explanations for this ethnic variation remain unclear.

A common genetic polymorphism of the G_i protein β_3 -subunit gene (*GNB3*) C⁸²⁵T is associated with hypertension, low renin levels, and obesity in some but not all populations of different ethnic backgrounds. In addition, the existence of an 825T allele predicted selective α_2 -AR-mediated coronary vasoconstriction. Although blacks have a higher allele frequency of 825T, the clinical significance is unknown.

In summary, proteins that determine drug disposition and response, such as drug-metabolizing enzymes, drug transporters, and drug receptors, are the products of the genes encoding them. The study of their molecular genetics may provide a clearer understanding of ethnic differences in drug response. The principal focus so far has been on drug metabolism and drug receptors. Future directions include

(a) characterizing the in vivo functional significance of known polymorphisms, (b) comparing phenotypic and genotypic characterization among multiple ethnic groups, (c) identifying new polymorphisms in genes that regulate drug disposition or response, and (d) defining the relative contribution of genetic and environmental factors to ethnic variations in drug response.

ACKNOWLEDGMENTS

Dr. Xie was a Merck International Fellow in Clinical Pharmacology. This work was supported by grants HL 56251, HL 04012, and GM 31304 from the US Public Health Service.

Visit the Annual Reviews home page at www.AnnualReviews.org

LITERATURE CITED

1. Senior PA, Bhopal R. 1994. Ethnicity as a variable in epidemiology research. *Br. Med. J.* 309:327–30
2. Editorial. 2000. Census, race and science. *Nature Genet.* 24:97–98
3. Lewis P, Rack PH, Vaddadi KS, Allen JJ. 1980. Ethnic differences in drug response. *Postgrad. Med. J.* 56(suppl.1): 46–49
4. Wood AJJ, Zhou HH. 1991. Ethnic differences in drug disposition and responsiveness. *Clin. Pharmacokinet.* 20:350–73
5. Smith MW, Mendoza RP. 1996. Ethnicity and pharmacogenetics. *Mt. Sinai J. Med.* 63:285–90
6. Wood AJJ. 1998. Ethnic differences in drug disposition and response. *Ther. Drug Monit.* 20:525–26
7. Johnson JA. 2000. Predictability of the effects of race or ethnicity on pharmacokinetics of drugs. *Intl. J. Clin. Pharmacol. Ther.* 38:53–60
8. Gaedigk A. 2000. Interethnic differences of drug-metabolizing enzymes. *Intl. J. Clin. Pharmacol. Ther.* 38:61–68
- 8a. Stein CM, Lang CC, Xie HG, Wood AJJ. 2001. Hypertension in blacks: Study of specific genotypes and phenotypes will provide a greater understanding of interindividual and interethnic variability in blood pressure regulation than studies based on race. *Pharmacogenetics* In press
9. Kalow W. 1982. Ethnic differences in drug metabolism. *Clin. Pharmacokinet.* 7:373–400
10. Kalow W. 1991. Interethnic variation of drug metabolism. *Trends Pharmacol. Sci.* 12:102–7
11. Kalow W, Bertilsson L. 1994. Interethnic factors affecting drug response. *Adv. Drug Res.* 25:1–53
12. Bertilsson L. 1995. Geographical/interracial differences in polymorphic drug oxidation: current state of knowledge of cytochromes P450 (CYP) 2D6 and 2C19. *Clin. Pharmacokinet.* 29:192–209
13. Meyer UA, Zanger UM. 1997. Molecular mechanisms of genetic polymorphisms of drug metabolism. *Annu. Rev. Pharmacol. Toxicol.* 37:269–96
14. Kalow W. 1984. Pharmacoanthropology: outline, problems, and the nature of case histories. *Fed. Proc.* 43:2134–38
15. Shimada T, Yamazaki H, Mimura M, Inui Y, Guengerich FP. 1994. Interindividual variations in human liver cytochrome P-450 enzymes involved in the oxidation of drugs, carcinogens and toxic chemicals: studies with liver microsomes of 30 Japanese and 30 Caucasians. *J. Pharmacol. Exp. Ther.* 270:414–23
16. Goldstein JA, de Morais SMF. 1994.

- Biochemistry and molecular biology of the human CYP2C subfamily. *Pharmacogenetics* 4:285–99
17. Nelson DR, Koymans L, Kamataki T, Stegeman JJ, Feyereisen R, et al. 1996. P-450 superfamily: update on the new sequences, gene mapping, accession numbers, and nomenclature. *Pharmacogenetics* 6:1–42
 18. Inoue K, Yamazaki H, Imiya K, Akasaka S, Guengerich FP, et al. 1997. Relationship between *CYP2C9* and *2C19* genotypes and tolbutamide methyl hydroxylation and *S*-mephenytoin 4'-hydroxylation activities in livers of Japanese and Caucasian populations. *Pharmacogenetics* 7:103–13
 19. Miners JO, Birkett DJ. 1998. Cytochrome P4502C9: an enzyme of major importance in human drug metabolism. *Br. J. Clin. Pharmacol.* 45:525–38
 20. Scott J, Poffenbarger PL. 1979. Pharmacogenetics of tolbutamide metabolism in humans. *Diabetes* 28:41–51
 21. Veronese ME, Miners JO, Rees DLP, Birkett DJ. 1993. Tolbutamide hydroxylation in humans: lack of bimodality in 106 healthy subjects. *Pharmacogenetics* 3:86–93
 22. Page MA, Boutagy JS, Shenfield GM. 1991. A screening test for slow metabolizers of tolbutamide. *Br. J. Clin. Pharmacol.* 31:649–54
 23. Spielberg S, McCrea J, Cribb A, Rushmore T, Waldman S, et al. 1996. A mutation in *CYP2C9* is responsible for decreased metabolism of losartan. *Clin. Pharmacol. Ther.* 59:215 (Abstr.)
 24. Sullivan-Klose TH, Ghanayem BI, Bell DA, Zhang ZY, Kaminsky LS, et al. 1996. The role of the *CYP2C9*-Leu359 allelic variant in the tolbutamide polymorphism. *Pharmacogenetics* 6:341–49
 25. Steward DJ, Haining RL, Henne KR, Davis G, Rushmore TH, et al. 1997. Genetic association between sensitivity to warfarin and expression of *CYP2C9**3. *Pharmacogenetics* 7:361–67
 26. Aynacioglu AS, Brockmüller J, Bauer S, Sachse C, Güzelbey P, et al. 1999. Frequency of cytochrome P450 *CYP2C9* variants in a Turkish population and functional relevance for phenytoin. *Br. J. Clin. Pharmacol.* 48:409–15
 27. McCrea JB, Cribb A, Rushmore T, Osborne B, Gillen L, et al. 1999. Phenotypic and genotypic investigations of a healthy volunteer deficient in the conversion of losartan to its active metabolite E-3174. *Clin. Pharmacol. Ther.* 65:348–52
 28. Aithal GP, Day CP, Kesteven PJJ, Daly AK. 1999. Association of polymorphisms in the cytochrome P450 *CYP2C9* with warfarin dose requirement and risk of bleeding complications. *Lancet* 353:717–19
 - 28a. Ogg MS, Brennan P, Meade T, Humphries SE. 1999. *CYP2C**3 allelic variants and bleeding complications. *Lancet* 354:1124
 29. Aithal GP, Day CP, Kesteven PJJ, Daly AK. 1999. Warfarin dose requirement and *CYP2C9* polymorphisms. *Lancet* 353:1972–73
 30. Meehan RR, Gosden JR, Rout D, Hasle ND, Friedberg T. 1988. Human cytochrome P450 PB-1: a multigene family involved in mephenytoin and steroid oxidations that maps to chromosome 10. *Am. J. Hum. Genet.* 42:26–37
 31. de Morais SMF, Schweikl H, Blaisdell J, Goldstein JA. 1993. Gene structure and upstream regulatory regions of human *CYP2C9* and *CYP2C18*. *Biochem. Biophys. Res. Commun.* 194:194–201
 32. Rettie AE, Wienkers LC, Gonzalez FJ, Trager WF, Korzekwa KR. 1994. Impaired (*S*)-warfarin metabolism catalyzed by the *R144C* allelic variant of *CYP2C9*. *Pharmacogenetics* 4:39–42
 33. Haining RL, Hunter AP, Veronese ME, Trager WF, Rettie AE. 1996. Allelic variants of human cytochrome P450 2C9: baculovirus-mediated expression, purification, structural characterization, substrate stereoselectivity, and prochiral

- selectivity of the wild-type and I359L mutant forms. *Arch. Biochem. Biophys.* 333:447–58
34. Miners JO, Coulter S, Birkett DJ, Goldstein JA. 2000. Torsemide metabolism by CYP2C9 variants and other human CYP2C subfamily enzymes. *Pharmacogenetics* 10:267–70
35. Takanashi K, Tainaka H, Kobayashi K, Yasumori T, Hosakawa M, et al. 2000. CYP2C Ile³⁵⁹ and Leu³⁵⁹ variants: enzyme kinetic study with seven substrates. *Pharmacogenetics* 10:95–104
36. Yamazaki H, Inoue K, Shimada T. 1998. Roles of two allelic variants (Arg144Cys and Ile359Leu) of cytochrome P450C9 in the oxidation of tolbutamide and warfarin by human liver microsomes. *Xenobiotica* 28:103–15
37. Kaminsky LS, de Morais SMF, Faletto MB, Dunbar D, Goldstein JA. 1993. Correlation of human cytochrome P450C substrate specificities with primary structure: warfarin as a probe. *Mol. Pharmacol.* 43:234–39
38. Furuya H, Fernandez-Salguero P, Gregory W, Taber H, Steward A, et al. 1995. Genetic polymorphism of CYP2C9 and its effect on warfarin maintenance dose requirement in patients undergoing anticoagulation therapy. *Pharmacogenetics* 5:389–92
39. Bhasker CR, Miners JO, Coulter S, Birkett DJ. 1997. Allelic and functional variability of cytochrome P450C9. *Pharmacogenetics* 7:51–58
40. Kidd RS, Straughn AB, Meyer MC, Blaisdell J, Goldstein JA, et al. 1999. Pharmacokinetics of chlorpheniramine, phenytoin, glipizide and nifedipine in an individual homozygous for the CYP2C9*3 allele. *Pharmacogenetics* 9:71–80
41. Miners J, Wing LMH, Birkett DJ. 1985. Normal metabolism of debrisoquine and theophylline in a slow tolbutamide metabolizer. *Aust. N.Z. J. Med.* 15:348–49
42. Mamiya K, Ieiri I, Shimamoto J, Yukawa E, Imai J, et al. 1998. The effects of genetic polymorphisms of CYP2C9 and CYP2C19 on phenytoin metabolism in Japanese adult patients with epilepsy: studies in stereoselective hydroxylation and population pharmacokinetics. *Epilepsia* 39:1317–23
43. Odani A, Hashimoto Y, Otsuki Y, Uwai Y, Hattori H, et al. 1997. Genetic polymorphism of the CYP2C subfamily and its effect on the pharmacokinetics of phenytoin in Japanese patients with epilepsy. *Clin. Pharmacol. Ther.* 62:287–92
44. Takahashi H, Kashima T, Nomizo Y, Muramoto N, Shimizu T, et al. 1998. Metabolism of warfarin enantiomers in Japanese patients with heart disease having different CYP2C9 and CYP2C19 genotypes. *Clin. Pharmacol. Ther.* 63:519–28
45. Takahashi H, Kashima T, Nomoto S, Iwade K, Tainaka H, et al. 1998. Comparisons between in-vitro and in-vivo metabolism of (S)-warfarin: catalytic activities of cDNA-expressed CYP2C9, its Leu359 variant and their mixture versus unbound clearance in patients with the corresponding CYP2C9 genotypes. *Pharmacogenetics* 8:365–73
- 45a. Thijssen HHW, Verkooyen IWC, Frank HLL. 2000. The possession of the CYP2C9*3 allele is associated with low dose requirement of acenocoumarol. *Pharmacogenetics* 10:757–60
46. Arnold K, Gerber N. 1970. The rate of decline of diphenylhydantoin in human plasma. *Clin. Pharmacol. Ther.* 11:121–34
47. Edeki TI, Brase DA. 1995. Phenytoin disposition and toxicity: role of pharmacokinetic and interethnic factors. *Drug Metab. Rev.* 27:449–69
- 47a. Yoon YR, Shon JH, Kim MK, Lim YC, Lee HR, et al. 2001. Frequency of cytochrome P450 2C9 mutant alleles in a Korean population. *Br. J. Clin. Pharmacol.* In press
48. Wang S-L, Huang J-D, Lai M-D, Tsai J-J.

1995. Detection of CYP2C9 polymorphism based on the polymerase chain reaction in Chinese. *Pharmacogenetics* 5:37–42
49. Nasu K, Kubota T, Ishizaki T. 1997. Genetic analysis of CYP2C9 polymorphism in a Japanese population. *Pharmacogenetics* 7:405–9
50. Kimura M, Ieiri I, Mamiya K, Urae A, Higuchi S. 1998. Genetic polymorphism of cytochrome P450s, CYP2C19, and CYP2C9 in a Japanese population. *Ther. Drug Monit.* 20:243–47
51. London SJ, Daly AK, Leathart JB, Navidi WC, Idle JR. 1996. Lung cancer risk in relation to the CYP2C9*1/CYP2C9*2 genetic polymorphism among African-Americans and Caucasians in Los Angeles county, California. *Pharmacogenetics* 6:527–33
52. Stubbins MJ, Harries LW, Smith G, Tarbit MH, Wolf CR. 1996. Genetic analysis of the human cytochrome P450 CYP2C9 locus. *Pharmacogenetics* 6:429–39
53. Brochmüller J, Rost KL, Gross D, Schenkel A, Roots I. 1995. Phenotyping of CYP2C19 with enantiospecific HPLC-quantification of R- and S-mephenytoin and comparison with the intron 4/exon 5 G→A-splice site mutation. *Pharmacogenetics* 5:80–88
54. Ackermann E, Cascorbi I, Sachse C, Brockmüller J, Mrozikiewicz PM, et al. 1997. Frequencies and the allelic linkage of CYP2C9 mutations in a German population, and the detection of a C/T mutation in intron 2. *Eur. J. Clin. Pharmacol.* 52:A71 (Abstr.)
55. Yasar Ü, Eliasson E, Dahl ML, Johansson I, Ingelman-Sundberg M, et al. 1999. Validation of methods for CYP2C9 genotyping: frequencies of mutant alleles in a Swedish population. *Biochem. Biophys. Res. Commun.* 254:628–31
56. Blann A, Hewitt J, Siddiqui F, Bareford D. 1999. Racial background is a determinant of average warfarin dose required to maintain the INR between 2.0 and 3.0. *Br. J. Haematol.* 107:207–9
57. Yu HCM, Chan TYK, Critchley JAJH, Woo KS. 1996. Factors determining the maintenance dose of warfarin in Chinese patients. *Q. J. Med.* 89:127–35
58. Schwarz UI, Choo EF, Dresser GK, Stein CM, Wood AJJ, et al. 2000. Identification of a new CYP2C9 variant in African-Americans. *Clin. Pharmacol. Ther.* 67:169 (Abstr.)
59. Imai J, Ieiri I, Mamiya K, Miyahara S, Furuumi H, et al. 2000. Polymorphism of the cytochrome P450 (CYP) 2C9 gene in Japanese epileptic patients: genetic analysis of the CYP2C9 locus. *Pharmacogenetics* 10:85–89
60. Gotoh O. 1992. Substrate recognition sites in cytochrome P-450 family 2 (CYP2) proteins inferred from comparative analysis of amino acid and coding nucleotide sequences. *J. Biol. Chem.* 267:83–90
61. Chang M, Tybring G, Dahl M-L, Götharson E, Sagar M, et al. 1995. Interphenotype differences in disposition and effect on gastrin levels of omeprazole - suitability of omeprazole as a probe for CYP2C19. *Br. J. Clin. Pharmacol.* 39:511–18
62. Caraco Y, Lagerstrom PO, Wood AJJ. 1996. Ethnic and genetic determinants of omeprazole disposition and effect. *Clin. Pharmacol. Ther.* 60:157–67
63. Furuta T, Ohashi K, Kamata T, Takashima M, Kosuge K, Kawasaki T, et al. 1998. Effect of genetic differences in omeprazole metabolism on cure rates for *Helicobacter pylori* infection and peptic ulcer. *Ann. Intern. Med.* 129:1027–30
- 63a. Ohashi K, Furuta T, Kosuge K, Kimura M, Nishimoto M, et al. 1998. Disposition and effects of omeprazole and related to CYP2C19 genotype. *Clin. Pharmacol. Ther.* 63:152 (Abstr.)
64. Furuta T, Ohashi K, Kosuge K, Zhao X-J, Takashima M, et al. 1999. CYP2C19

- genotype status and effect of omeprazole on intragastric pH in humans. *Clin. Pharmacol. Ther.* 65:552–61
65. Furuta T, Takashima M, Shirai N, Xiao F, Hanai H, et al. 2000. Cure of refractory duodenal ulcer and infection caused by *Helicobacter pylori* by high doses of omeprazole and amoxicillin in a homozygous CYP2C19 extensive metabolizer patient. *Clin. Pharmacol. Ther.* 67:684–89
66. Tanigawara Y, Aoyama N, Kita T, Shirakawa K, Komada F, Kasuga M, et al., 1999. CYP2C19 genotype-related efficacy of omeprazole for the treatment of infection caused by *Helicobacter pylori*. *Clin. Pharmacol. Ther.* 66:528–34
67. Daly AK. 1995. Molecular basis of polymorphic drug metabolism. *J. Mol. Med.* 73:539–53
68. Romkes M, Faletto MB, Blaisdell JA, Raucy JL, Goldstein JA. 1991. Cloning and expression of complementary DNA for multiple members of the human cytochrome P450 IIC subfamily. *Biochemistry* 30:3247–55
69. Zaphiropoulos PG. 1999. RNA molecules containing exons originating from different members of the cytochrome P450 2C gene subfamily (CYP2C) in human epidermis and liver. *Nucleic Acids Res.* 27:2585–90
70. de Morais SMF, Wilkinson GR, Blaisdell J, Nakamura K, Meyer UA, et al. 1994. The major genetic defect responsible for the polymorphism of *S*-mephenytoin metabolism in humans. *J. Biol. Chem.* 269:15419–22
71. de Morais SMF, Wilkinson GR, Blaisdell J, Meyer UA, Nakamura K, et al. 1994. Identification of a new genetic defect responsible for the polymorphism of (S)-mephenytoin metabolism in Japanese. *Mol. Pharmacol.* 46:594–98
72. de Morais SMF, Goldstein JA, Xie HG, Huang SL, Lu YQ, et al. 1995. Genetic analysis of the *S*-mephenytoin in a Chinese population. *Clin. Pharmacol. Ther.* 58:404–11
73. Xie HG, Stein CM, Kim RB, Wilkinson GR, Flockhart DA, et al. 1999. Allelic, genotypic, and phenotypic distribution of *S*-mephenytoin 4'-hydroxylase (CYP2C19) in healthy Caucasian populations of European descent throughout the world. *Pharmacogenetics* 9:539–49
74. Xie HG, Kim RB, Stein CM, Wilkinson GR, Wood AJJ. 1999. Genetic polymorphism of (S)-mephenytoin 4'-hydroxylation in populations of African descent. *Br. J. Clin. Pharmacol.* 48:402–8
75. Xie HG. 2000. Genetic variations of *S*-mephenytoin 4'-hydroxylase (CYP2C19) in the Chinese population. *Life Sci.* 66:PL175–81
76. Alván G, Bechtel P, Iselius L, Gundert-Remy U. 1990. Hydroxylation polymorphisms of debrisoquine and mephenytoin in European populations. *Eur. J. Clin. Pharmacol.* 39:533–37
77. Wilkinson GR, Guengerich FP, Branch RA. 1989. Genetic polymorphism of *S*-mephenytoin hydroxylation. *Pharmacol. Rev.* 43:53–76
78. Zhang Y, Reviriego J, Lou YQ, Sjöqvist F, Bertilsson L. 1990. Diazepam metabolism in native Chinese poor and extensive hydroxylators of *S*-mephenytoin: interethnic differences in comparison with white subjects. *Clin. Pharmacol. Ther.* 48:496–502
79. Bertilsson L, Henthorn TK, Sanz E, Tybring G, Säwe J, et al. 1989. Importance of genetic factors in the regulation of diazepam metabolism: relationship to *S*-mephenytoin, but not debrisoquin, hydroxylation phenotype. *Clin. Pharmacol. Ther.* 45:348–55
80. Bertilsson L, Kalow W. 1993. Why are diazepam metabolism and polymorphic *S*-mephenytoin hydroxylation associated with each other in white and Korean populations but not in Chinese population? *Clin. Pharmacol. Ther.* 53:608–10
81. Xie HG. 1997. Direct evidence for the

- higher frequency of *CYP2C19* allelic heterozygotes in Chinese subjects than in white subjects. *Clin. Pharmacol. Ther.* 62:691–92
82. Kumana CR, Lauder IJ, Chan M, Ko W, Lin HJ. 1987. Differences in diazepam pharmacokinetics in Chinese and white Caucasians-relation to body lipid stores. *Eur. J. Clin. Pharmacol.* 32:211–15
83. Andersson T, Regårdh CG, Lou YC, Zhang Y, Dahl ML, et al. 1992. Polymorphic hydroxylation of *S*-mephenytoin and omeprazole metabolism in Caucasian and Chinese subjects. *Pharmacogenetics* 2:25–31
84. Caraco Y, Lagestrom PO, Wood AJJ. 1995. Omeprazole disposition in Caucasian and Chinese subjects. *Clin. Pharmacol. Ther.* 57:216 (Abstr.)
85. Caraco Y, Tateishi T, Wood AJJ. 1995. Interethnic difference in omeprazole's inhibition of diazepam metabolism. *Clin. Pharmacol. Ther.* 58:62–72
86. Ishizaki T, Sohn DR, Kobayashi K, Chiba K, Lee KH, et al. 1994. Interethnic differences in omeprazole metabolism in the two *S*-mephenytoin hydroxylation phenotype studied in Caucasians and Orientals. *Ther. Drug Monit.* 16:214–15
87. Funck-Brentano C, Becquemont L, Leneveu A, Roux A, Jaillon P, et al. 1997. Inhibition by omeprazole of proguanil metabolism: mechanism of the interaction *in vitro* and prediction of *in vivo* results from the *in vivo* experiments. *J. Pharmacol. Exp. Ther.* 280:730–38
88. Kaneko A, Bergqvist Y, Takechi M, Kalkoa M, Kaneko O, et al. 1999. Intrinsic efficacy of proguanil against falciparum and vivax malaria independent of the metabolite cycloguanil. *J. Infect. Dis.* 179:974–79
89. Idle JR, Corchero J, Gonzalez FJ. 2000. Medical implications of HGP's sequence of chromosome 22. *Lancet* 355:319
90. Xie HG, Xu ZH, Luo X, Huang SL, Zeng FD, et al. 1996. Genetic polymorphisms of debrisoquine and *S*-mephenytoin oxidation metabolism in Chinese populations: a meta-analysis. *Pharmacogenetics* 6:235–38
91. Lennard MS, Iyuan AO, Jackson PR, Tucker GT, Woods HF. 1992. Evidence for a disassociation in the control of sparteine, debrisoquine and metoprolol metabolism in Nigerians. *Pharmacogenetics* 2:89–92
92. Sachse C, Brockmüller J, Bauer S, Roots I. 1997. Cytochrome P450 2D6 variants in a Caucasian population: allele frequencies and phenotypic consequences. *Am J. Hum. Genet.* 60:284–95
93. Gaedigk A, Gotschall RR, Forbes NS, Simon SD, Kearns GL, et al. 1999. Optimization of cytochrome P4502D6 (*CYP2D6*) phenotype assignment using a genotyping algorithm based on allele frequency data. *Pharmacogenetics* 9:669–82
94. Wennerholm A, Johansson I, Massele AY, Jande M, Alm C, et al. 1999. Decreased capacity for debrisoquine metabolism among black Tanzanians: analyses of the *CYP2D6* genotype and phenotype. *Pharmacogenetics* 9:707–14
95. Fukuda T, Yamamoto I, Nishida Y, Zhou Q, Ohno M, et al. 1999. Effect of the *CYP2D6**10 genotype on venlafaxine pharmacokinetics in healthy adult volunteers. *Clin. Pharmacol. Ther.* 47:450–53
96. Johansson I, Oscarson M, Yue Q-Y, Bertilsson L, Sjöqvist F, et al. 1994. Genetic analysis of the Chinese cytochrome P4502D locus: characterization of variant *CYP2D6* genes present in subjects with diminished capacity for debrisoquine hydroxylation. *Mol. Pharmacol.* 46:452–59
97. Hou ZY, Chen CP, Yang WC, Lai MD, Buchert ET, et al. 1996. Determination of dextromethorphan metabolic phenotype by salivary analysis with a reference to genotype in Chinese patients receiving renal hemodialysis. *Clin. Pharmacol. Ther.* 59:411–17
98. Lai ML, Wang SL, Lai MD, Lin ET, Tse M, et al. 1995. Propranolol disposition in Chinese subjects of different *CYP2D6* genotypes. *Clin. Pharmacol. Ther.* 58:264–68

99. Oscarson M, Hidestrand M, Johansson I, Ingelman-Sundberg M. 1997. A combination of mutations in the *CYP2D6*17* (*CYP2D6Z*) allele causes alterations in enzyme function. *Mol. Pharmacol.* 52:1034–40
100. Tateishi T, Chida M, Ariyoshi N, Mizorogi Y, Kamataki T, et al. 1999. Analysis of the *CYP2D6* gene in relation to dextromethorphan *O*-demethylation capacity in a Japanese population. *Clin. Pharmacol. Ther.* 65:570–75
101. Tseng CY, Wang SL, Lai MD, Lai ML, Huang JD. 1996. Formation of morphine from codeine in Chinese subjects of different *CYP2D6* genotypes. *Clin. Pharmacol. Ther.* 60:177–82
102. Yokota H, Tamura S, Furuya H, Kimura S, Watanabe M, et al. 1993. Evidence for a new variant *CYP2D6* allele *CYP2D6J* in a Japanese population associated with lower in vivo rates of sparteine metabolism. *Pharmacogenetics* 3:256–63
103. Bertilsson L, Lou Y-Q, Du Y-L, Liu Y, Kuang T-Y, et al. 1992. Pronounced differences between native Chinese and Swedish populations in the polymorphic hydroxylations of debrisoquin and *S*-mephenytoin. *Clin. Pharmacol. Ther.* 51:388–97
104. Lou YC. 1990. Differences in drug metabolism polymorphism between Orientals and Caucasians. *Drug Metab. Rev.* 22:451–75
105. Rudorfer MV, Lane EA, Chang W-H, Zhang MD, Potter WZ. 1984. Desipramine pharmacokinetics in Chinese and Caucasian volunteers. *Br. J. Clin. Pharmacol.* 17:433–40
106. Yue QY, Svensson JO, Alm C, Sjöqvist F, Säwe J. 1989. Interindividual and interethnic differences in the demethylation and glucuronidation of codeine. *Br. J. Clin. Pharmacol.* 28:629–37
107. Yue QY, Svensson JO, Sjöqvist F, Säwe J. 1991. A comparison of the pharmacokinetics of codeine and its metabolites in healthy Chinese and Caucasian extensive hydroxylators of debrisoquine. *Br. J. Clin. Pharmacol.* 31:643–47
108. Caraco Y, Sheller J, Wood AJJ. 1999. Impact of ethnic origin and quinidine coadministration on codeine's disposition and pharmacodynamic effects. *J. Pharmacol. Exp. Ther.* 290:413–22
109. Horai Y, Nakano M, Ishizaki T, Ishikawa K, Zhou HH, et al. 1989. Metoprolol and mephenytoin oxidation polymorphism in Far Eastern Oriental subjects: Japanese versus mainland Chinese. *Clin. Pharmacol. Ther.* 46:198–207
110. Sohn DR, Shin SG, Park CW, Kusaka M, Chiba K, et al. 1991. Metoprolol oxidation polymorphism in a Korean population: comparison with native Japanese and Chinese populations. *Br. J. Clin. Pharmacol.* 32:504–7
111. Roh HK, Dahl ML, Johansson I, Ingelman-Sundberg M, Cha YN, et al. 1996. Debrisoquine and *S*-mephenytoin hydroxylation phenotypes and genotypes in a Korean population. *Pharmacogenetics* 6:441–47
112. Yue QY, Svensson JO, Säwe J, Bertilsson L. 1995. Codeine metabolism in three Oriental populations: a pilot study in Chinese, Japanese and Koreans. *Pharmacogenetics* 5:173–77
- 112a. Nishida Y, Fukuda T, Yamamoto I, Azuma J. 2000. *CYP2D6* genotypes in a Japanese population: low frequencies of *CYP2D6* gene duplication but high frequency of *CYP2D6*10*. *Pharmacogenetics* 10:567–70
113. Wang SL, Huang JD, Lai MD, Liu BH, Lai ML. 1993. Molecular basis of genetic variation in debrisoquine hydroxylation in Chinese subjects: polymorphism in RFLP and DNA sequence of *CYP2D6*. *Clin. Pharmacol. Ther.* 53:410–18

114. Garcia-Barceló M, Chow LY, Kum Chiu HF, Wing YK, Shing Lee DT, et al. 2000. Genetic analysis of the *CYP2D6* locus in a Hong Kong Chinese population. *Clin. Chem.* 46:18–23
115. Kubota T, Yamaura Y, Ohkawa N, Hara H, Chiba K. 2000. Frequencies of *CYP2D6* mutant alleles in a normal Japanese population and metabolic activity of dextromethorphan *O*-demethylation in different *CYP2D6* genotypes. *Br. J. Clin. Pharmacol.* 50:31–34
116. Forbes NS, Bradford LD, Gotschall RR, Leeder JS, Gaedigk A. 1999. *CYP2D6* allele frequencies in African Americans: phenotype concordance with dextromethorphan. *Clin. Pharmacol. Ther.* 65:170 (Abstr.)
117. Leathart JBS, London SJ, Steward A, Adams JD, Idle JR. et al. 1998. *CYP2D6* phenotype-genotype relationships in African-Americans and Caucasians in Los Angeles. *Pharmacogenetics* 8:529–41
118. London SJ, Daly AK, Leathart JBS, Navidi WC, Carpenter CC, et al. 1997. Genetic polymorphism of *CYP2D6* and lung cancer risk in African-Americans and Caucasians in Los Angeles county. *Carcinogenesis* 18:1203–14
119. Martin DE, Tran JQ, Flockhart DA, Jorkasky DK. 1998. Analysis of *CYP2D6* and *CYP2C19* genotypes in large African-American and Caucasian populations. *Clin. Pharmacol. Ther.* 63:206 (Abstr.)
120. Evans WE, Relling MV, Rahman A, McLeod HL, Scott EP, et al. 1993. Genetic basis for a lower prevalence of deficient *CYP2D6* oxidative drug metabolism phenotypes in black Americans. *J. Clin. Invest.* 91:2150–54
121. Aklillu E, Persson I, Bertilsson L, Johansson I, Rodrigues F, et al. 1996. Frequent distribution of ultrarapid metabolizers of debrisoquine in an Ethiopian population carrying duplicated and multiplicated functional *CYP2D6* alleles. *J. Pharmacol. Exp. Ther.* 278:441–46
122. Bathum L, Skjelbo E, Mutabingwa TK, Madsen H, Hørder M, et al. 1999. Phenotypes and genotypes for *CYP2D6* and *CYP2C19* in a black Tanzanian population. *Br. J. Clin. Pharmacol.* 48:395–401
123. Masimirembwa C, Hasler J, Bertilsson L, Johansson I, Ekberg O, et al. 1996. Phenotype and genotype analysis of debrisoquine hydroxylase (*CYP2D6*) in a black Zimbabwean population. Reduced enzyme activity and evaluation of metabolic correlation of *CYP2D6* probe drugs. *Eur. J. Clin. Pharmacol.* 51:117–22
124. Masimirembwa C, Persson I, Bertilsson L, Hasler J, Ingelman-Sundberg M. 1996. A novel mutant variant of the *CYP2D6* gene (*CYP2D6*17*) common in a black African population: association with diminished debrisoquine hydroxylase activity. *Br. J. Clin. Pharmacol.* 42:713–19
125. Chen S, Chou W-H, Blouin RA, Mao Z, Humphries LL, et al. 1997. The cytochrome P450 2D6 (*CYP2D6*) enzyme polymorphism: screening costs and influence on clinical outcomes in psychiatry. *Clin. Pharmacol. Ther.* 60:522–34
126. Marez D, Legrand M, Sabbagh N, Guidice JM, Spire C, et al. 1997. Polymorphism of the cytochrome P450 *CYP2D6* gene in European population: characterization of 48 mutations and 53 alleles, their frequencies and evolution. *Pharmacogenetics* 7:193–202
127. Madsen H, Nielsen KK, Brøsen K. 1995. Imipramine metabolism in relation to the sparteine and mephenytoin oxidation polymorphisms—a population study. *Br. J. Clin. Pharmacol.* 39:433–39
128. Bathum L, Andersen-Ranberg K, Boldsen J, Brøsen K, Jeune B. 1998. Genotypes for the cytochrome P450 enzymes *CYP2D6* and *CYP2C19* in human longevity: role of *CYP2D6* and *CYP2C19* in longevity. *Eur. J. Clin. Pharmacol.* 54:427–30
129. Marandi T, Dahl ML, Kiivet RA, Rägo L,

- Sjöqvist F. 1996. Debrisoquine and S-mephenytoin hydroxylation phenotypes and CYP2D6 genotypes in an Estonian population. *Pharmacol. Toxicol.* 78:303–7
130. Saarikoski ST, Sata F, Husgafvel-Pursiainen K, Rautalahti M, Haukka J, et al. 2000. *CYP2D6* ultrarapid metabolizer genotype as a potential modifier of smoking behavior. *Pharmacogenetics* 10:5–10
131. Griese EU, Zanger UM, Brudermanns U, Gaedigk A, Mikus G, et al. 1998. Assessment of the predictive power of genotypes for the in vivo catalytic function of CYP2D6 in a Caucasian population. *Pharmacogenetics* 8:15–26
132. Marandi T, Dahl M-L, Rågo L, Kiviet R, Sjöqvist F. 1997. Debrisoquine and S-mephenytoin hydroxylation polymorphisms in a Russian population living in Estonia. *Eur. J. Clin. Pharmacol.* 53:257–60
133. Yamada H, Dahl M-L, Lannfelt L, Viitanen M, Winblad B, et al. 1998. *CYP2D6* and *CYP2C19* genotypes in an elderly Swedish population. *Eur. J. Clin. Pharmacol.* 54:479–81
134. Aynacioglu AS, Sachse C, Bozkurt A, Kortunay S, Nacak M, et al. 1999. Low frequency of defective alleles of cytochrome P450 enzymes 2C19 and 2D6 in the Turkish population. *Clin. Pharmacol. Ther.* 66:185–92
- 134a. Ingelman-Sundberg M, Oscarson M, McLellan RA. 1999. Polymorphic human cytochrome P450 enzymes: an opportunity for individualized drug treatment. *Trends Pharmacol. Sci.* 20: 342–49
135. Dahl M-L, Johansson I, Porsmyr-Palmertz M, Ingelman-Sundberg M. 1992. Analysis of the *CYP2D6* gene in relation to debrisoquine and desipramine hydroxylation in a Swedish population. *Clin. Pharmacol. Ther.* 51:12–17
136. Rendic S, Di Carlo FJ. 1997. Human cytochrome P450 enzymes: a status report summarizing their reactions, substrates, inducers, and inhibitors. *Drug Metab. Rev.* 29:413–580
137. Soons PA, Schellens JHM, Breimer DD. 1992. Variability in pharmacokinetics and metabolism of nifedipine and dihydropyridine calcium entry blockers. In *Pharmacogenetics of Drug Metabolism*, ed. W Kalow, pp. 769–89. New York: Pergamon
138. Watkins PB. 1994. Non-invasive tests of CYP3A enzymes. *Pharmacogenetics* 4:171–84
139. Wachter VJ, Wu CY, Benet LZ. 1995. Overlapping substrate specificities and tissue distribution of cytochrome P450 3A and P-glycoprotein: implications for drug delivery and activity in cancer therapy. *Mol. Carcinog.* 13:129–34
140. Guengerich FP. 1999. Cytochrome P-450 3A4: Regulation and role in drug metabolism. *Annu. Rev. Pharmacol. Toxicol.* 39:1–17
141. Thummel KE, Wilkinson GR. 1998. In vitro and in vivo drug interactions involving human CYP3A. *Annu. Rev. Pharmacol. Toxicol.* 38:389–430
142. Guengerich FP, Martin MV, Beaune PH, Kremers P, Wolff T, et al., 1986. Characterization of rat and human liver microsomal cytochrome P-450 forms involved in nifedipine oxidation, a prototype for genetic polymorphism in oxidative drug metabolism. *J. Biol. Chem.* 261:5051–60
143. Kronbach T, Fischer V, Meyer U. 1988. Cyclosporine metabolism in human liver: identification of a cytochrome P450III gene family as the major cyclosporine-metabolizing enzyme explains interactions of cyclosporine with other drugs. *Clin. Pharmacol. Ther.* 43:630–35
144. Shimada T, Guengerich FP. 1989. Evidence for cytochrome P-450NF, the nifedipine oxidase, being the principal enzyme involved in the bioactivation of

- aflatoxins in human liver. *Proc. Natl. Acad. Sci. USA* 86:462–65
145. Lown KS, Thummel KE, Benedict PE, Shen DD, Turgeon DK, et al. 1995. The erythromycin breath test predicts the clearance of midazolam. *Clin. Pharmacol. Ther.* 57:16–24
146. Lindholm A, Henricsson S, Lind M, Dahlqvist R. 1988. Interindividual variability in the relative systemic availability of cyclosporine after oral dosing. *Eur. J. Clin. Pharmacol.* 34:461–64
147. Renwick AG, Robertson DRC, Macklin B, Challenor V, Waller DG, et al. 1988. The pharmacokinetics of oral nifedipine: a population study. *Br. J. Clin. Pharmacol.* 25:701–8
148. Schellens JHM, Soons PA, Breimer DD. 1988. Lack of bimodality in nifedipine plasma kinetics in a large population of healthy subjects. *Biochem. Pharmacol.* 37:2507–10
149. Thummel KE, Shen DD, Podoll TD, Kunze KL, Trager WF, et al. 1994. Use of midazolam as a human cytochrome P-450 3A probe: in vitro-in vivo correlation in liver transplant patients. *J. Pharmacol. Exp. Ther.* 271:549–56
150. Lin K-M, Lau JK, Smith R, Phillips P, Antal E, et al. 1988. Comparison of alprazolam plasma levels in normal Asian and Caucasian male volunteers. *Psychopharmacology* 96:365–69
151. Mück W, Unger S, Kawano K, Ahr G. 1998. Inter-ethnic comparisons of the pharmacokinetics of the HMG-CoA reductase inhibitor cerivastatin. *Br. J. Clin. Pharmacol.* 45:583–90
152. Stein CM, Sadeque AJ, Wandel C, Kim RB, Murray JJ, et al., 1999. Cyclosporine pharmacokinetics and pharmacodynamics in African-Americans and Caucasians. *Clin. Pharmacol. Ther.* 65:160 (Abstr.)
153. Lindholm A, Welsh M, Alton C, Kahan BD. 1992. Demographic factors influencing cyclosporine pharmacokinetic parameters in patients with uremia: racial differences in bioavailability. *Clin. Pharmacol. Ther.* 52:359–71
154. Schroeder TJ, Shah M, Hariharan S, First MR. 1996. Increased resources are required in patients with low cyclosporine bioavailability. *Transplant. Proc.* 28:2151–55
155. Yu KS, Bae KS, Sohn JH, Cho JY, Lee SY, et al. 2000. Ethnic differences in pharmacokinetics of nifedipine and erythromycin. *Clin. Pharmacol. Ther.* 67:161 (Abstr.)
156. Wandel C, Witte JS, Hall JM, Stein CM, Wood AJJ, et al. 2000. CYP3A activity in African American and European American men: population differences and functional effect of the *CYP3A4*1B* 5'-promoter region polymorphism. *Clin. Pharmacol. Ther.* 68:82–91
157. Ahsan CH, Renwick AG, Macklin B, Challenor VF, Waller DG, et al. 1991. Ethnic differences in the pharmacokinetics of oral nifedipine. *Br. J. Clin. Pharmacol.* 31:399–403
158. Ahsan CH, Renwick AG, Waller DG, Challenor VF, George CF, et al. 1993. The influences of dose and ethnic origins on the pharmacokinetics of nifedipine. *Clin. Pharmacol. Ther.* 54:329–38
159. Rashid TJ, Martin U, Clarke H, Waller DG, Renwick AG, et al. 1995. Factors affecting the absolute bioavailability of nifedipine. *Br. J. Clin. Pharmacol.* 40:51–58
160. Mohamed Z, Mustafa AM, Jamal SK, Mustafa MR, Jeyalingam K, et al. 1998. Cytochrome P-450 3A4 activity in two ethnic groups in Malaysia. *Clin. Pharmacol. Ther.* 63:221 (Abstr.)
161. Beerahee M, Wilkins MR, Jack DB, Beevers DG, Kendall MJ. 1987. Twelve hour (trough) plasma nifedipine concentrations during chronic treatment with nifedipine retard. *Eur. J. Clin. Pharmacol.* 32:347–49
162. Sowunmi A, Rashid TJ, Akinyinka OO, Renwick AG. 1995. Ethnic differences in

- nifedipine kinetics: comparisons between Nigerians, Caucasians and South Asians. *Br. J. Clin. Pharmacol.* 40:489–93
163. Kinirons MT, Lang CC, He HB, Ghebreselasie K, Shay S, et al. 1996. Triazolam pharmacokinetics and pharmacodynamics in Caucasians and Southern Asians: ethnicity and CYP3A activity. *Br. J. Clin. Pharmacol.* 41:69–72
164. Ladona MG, Linderström B, Thyr C, Peng DR, Rane A. 1991. Differential foetal development of the *O*- and *N*-demethylation of codeine and dextromethorphan in man. *Br. J. Clin. Pharmacol.* 32:295–302
165. Kleinbloesem CH, van Brummelen P, Faber H, Danhof M, Vermeulen NPE, et al. 1984. Variability in nifedipine pharmacokinetics and dynamics: a new oxidation polymorphism in man. *Biochem. Pharmacol.* 33:3721–24
166. Horsmans Y, Desager JP, Harvengt C. 1992. Absence of CYP3A genetic polymorphism assessed by urinary excretion of 6 β -hydroxycortisol in 102 healthy subjects on rifampicin. *Pharmacology* 71:258–61
167. Brooks BA, McBride OW, Dolphin CT, Farrall M, Scambler PJ, et al. 1988. The gene CYP3 encoding P-450pcn1 (nifedipine oxidase) is tightly linked to the gene COL1A2 encoding collagen type 1 alpha on 7q21-q22.1. *Am. J. Hum. Genet.* 43:280–84
168. Gonzalez FJ, Schmid BJ, Umeno M, McBride OW, Hardwick JP, et al. 1988. Human P450PCN1: sequence, chromosome localization, and direct evidence through cDNA expression that P450PCN1 is nifedipine oxidase. *DNA* 7:79–86
169. Spurr NK, Gough AC, Stevenson K, Wolf CR. 1989. The human cytochrome P-450 CYP3A locus: assignment to chromosome 7q22-qter. *Hum. Genet.* 81:171–74
170. Inoue K, Inazawa J, Nakagawa H, Shimada T, Yamazaki H, et al., 1992. Assignment of the human cytochrome P-450 nifedipine oxidase gene (CYP3A4) to chromosome 7 at band q22.1 by fluorescence in situ hybridization. *Jpn. J. Hum. Genet.* 37:133–38
171. Scherer SW, Rommens JM, Soder S, Wong E, Plavsic N, et al. 1993. Refined localization and yeast artificial chromosome (YAC) contig-mapping of genes and DNA segments in the 7q21-q32 region. *Hum. Mol. Genet.* 2:751–60
172. Hashimoto H, Toide K, Kitamura R, Fujita M, Tagawa S, et al. 1993. Gene structure of CYP3A4, an adult-specific form of cytochrome P450 in human livers, and its transcriptional control. *Eur. J. Biochem.* 218:585–95
173. Molowa DT, Schuetz EG, Wrighton SA, Watkins PB, Kremers P, et al. 1986. Complete cDNA sequence of a cytochrome P450 inducible by glucocorticoids in human liver. *Proc. Natl. Acad. Sci. USA* 83:5311–15
174. Beaune PH, Umbenhauer DR, Bork RW, Lloyd RS, Guengerich FP. 1986. Isolation and sequence determination of a cDNA clone related to human cytochrome P450 nifedipine oxidase. *Proc. Natl. Acad. Sci. USA* 83:8064–68
175. Bork RW, Muto T, Beaune PH, Srivastava PK, Lloyd RS, et al. 1989. Characterization of mRNA species related to human liver cytochrome P-450 nifedipine oxidase and regulation of catalytic activity. *J. Biol. Chem.* 264:910–19
176. Jounaïdi Y, Guzelian PS, Vilarem MJ. 1994. Sequence of the 5'-flanking region of CYP3A5: comparative analysis with CYP3A4 and CYP3A7. *Biochem. Biophys. Res. Commun.* 205:1741–47
177. Rebbeck TR, Jaffe JM, Walker AH, Wein AJ, Malkowicz SB. 1998. Modification of clinical presentation of prostate tumors by a novel genetic variant in CYP3A4. *J. Natl. Cancer Inst.* 90:1225–29
178. Felix CA, Walker AH, Lange BJ,

- Williams TM, Winick NJ, et al. 1998. Association of CYP3A4 genotype with treatment-related leukemia. *Proc. Natl. Acad. Sci. USA* 95:13176–81
179. Ball SE, Scatina J, Kao J, Ferron GM, Fruncillo R, et al. 1999. Population distribution and effects on drug metabolism of a genetic variants in the 5' promoter region of CYP3A4. *Clin. Pharmacol. Ther.* 66:288–94
180. Westlind A, Löfberg L, Tindberg N, Andersson TB, Ingelman-Sundberg M. 1999. Interindividual differences in hepatic expression of CYP3A4: relationship to genetic polymorphism in the 5'-upstream regulatory region. *Biochem. Biophys. Res. Commun.* 259:201–5
181. von Moltke LL, Tran TH, Cotreau MM, Greenblatt DJ. 2000. Unusually low clearance of two CYP3A substrates, alprazolam and trazodone, in a volunteer subject with wild-type CYP3A4 promoter region. *J. Clin. Pharmacol.* 40:200–4
- 181a. Tayeb MT, Clark C, Ameyaw M-M, Haites NE, Price Evans DA, et al. 2000. CYP3A4 promoter variant in Saudi, Ghanaian and Scottish Caucasian populations. *Pharmacogenetics* 10:753–56
182. Walker AH, Jaffe JM, Gunasegaram S, Cummings SA, Huang CS, et al. 1998. Characterization of an allelic variant in the nifedipine-specific element of CYP3A4: ethnic distribution and implications for prostate cancer risk. *Hum. Mutat.* 12:289
183. Sata F, Sapone A, Elizondo G, Stocker P, Miller VP, et al. 2000. CYP3A4 allelic variants with amino acid substitutions in exons 7 and 12: Evidence for an allelic variant with altered catalytic activity. *Clin. Pharmacol. Ther.* 67:48–56
184. Kim RB. 2000. Transporters and drug disposition. *Curr. Opin. Drug Dis. Dev.* 3:94–101
185. Yu DK. 1999. The contribution of P-glycoprotein to pharmacokinetic drug-drug interactions. *J. Clin. Pharmacol.* 39:1203–11
186. Rodriguez I, Abernethy DR, Woosley RL. 1999. P-glycoprotein in clinical cardiology. *Circulation* 99:472–74
187. Fromm MF. 2000. P-glycoprotein: a defense mechanism limiting oral bioavailability and CNS accumulation of drugs. *Int. J. Clin. Pharmacol. Ther.* 38:69–74
188. Hoffmeyer S, Burk O, von Richter O, Arnold HP, Brockmöller J, et al. 2000. Functional polymorphisms of the human multidrug-resistance gene: multiple sequence variations and correlation of one allele with P-glycoprotein expression and activity in vivo. *Proc. Natl. Acad. Sci. USA* 97:3473–78
189. Cordon-Cardo C, O'Brien JP, Boccia J, Casals D, Bertino JR, et al. 1990. Expression of the multidrug resistance gene product (P-glycoprotein) in human normal and tumor tissues. *J. Histochem. Cytochem.* 38:1277–87
190. Lown KS, Mayo RR, Leichtman AB, Hsiao H-L, Turgeon DK, et al. 1997. Role of intestinal P-glycoprotein (*mdr1*) in interpatient variation in the oral bioavailability of cyclosporine. *Clin. Pharmacol. Ther.* 62:248–60
191. Masuda S, Uemoto S, Hashida T, Inomata Y, Tanaka K, et al. 2000. Effect of intestinal P-glycoprotein on daily tacrolimus trough level in a living-donor small bowel recipient. *Clin. Pharmacol. Ther.* 68:98–103
192. Lindholm A, Kahan BD. 1993. Influence of cyclosporine pharmacokinetics, trough concentrations, and AUC monitoring on outcome after kidney transplantation. *Clin. Pharmacol. Ther.* 54:205–18
193. Schroeder TJ, Hariharan S, First MR. 1995. Variations in bioavailability of cyclosporine and relationship to clinical outcome in renal transplant subpopulations. *Transplant. Proc.* 27:837–39
194. Andrews PA, Sen M, Chang RWS. 1996.

- Racial variation in dosage requirements of tacrolimus. *Lancet* 348:1446
195. Choo EF, McKinsey J, Aresser GK, Mayo G, Kim RB, et al. 2000. Fexofenadine disposition is similar between Caucasian and African-American men and women. *Clin. Pharmacol. Ther.* 67:108 (Abstr.)
196. Gottesman MM, Hrycyna CA, Schoenlein PV, Germann UA, Pastan I. 1995. Genetic analysis of the multidrug transporter. *Annu. Rev. Genet.* 29:607–47
197. Ambudkar SV, Dey S, Hrycyna CA, Ramachandra M, Pastan I, et al. 1999. Biochemical, cellular, and pharmacological aspects of the multidrug transporter. *Annu. Rev. Pharmacol. Toxicol.* 39:361–98
198. Mickley LA, Lee J-S, Weng Z, Zhan Z, Alvarez M, et al. 1998. Genetic polymorphism in *MDR-1*: a tool for examining allelic expression in normal cells, unselected and drug-selected cell lines, and human tumors. *Blood* 91:1749–56
- 198a. Kim RB, Leake B, Choo E, Dresser GK, Kubba SV, et al. 2000. Identification of functionally important *MDR1* variant alleles among African-American and Caucasian subjects. *Drug Metab. Rev.* 32(suppl. 2):199 (Abstr.)
199. Hieble JP, Bylund DB, Clarke DE, Eikenburg DC, Langer SZ, et al. 1995. International Union of Pharmacology X. Recommendation for nomenclature of α_1 -adrenoceptors: Consensus Update. *Pharmacol. Rev.* 47:267–70
200. Brodde O-E, Michel MC. 1999. Adrenergic and muscarinic receptors in the human heart. *Pharmacol. Rev.* 51:651–89
201. Rudner XL, Berkowitz DE, Booth JV, Funk BL, Cozart KL, et al. 1999. Subtype specific regulation of human vascular α_1 -adrenergic receptor by vessel bed and age. *Circulation* 100:2336–43
202. Ricci A, Bronzetti E, Conterno A, Greco S, Mulatero P, et al. 1999. α_1 -Adrenergic receptor subtypes in human peripheral blood lymphocytes. *Hypertension* 33:708–12
203. Dimsdale JE, Graham RM, Ziegler MG, Zusman RM, Berry CC. 1987. Age, race, diagnosis, and sodium effects on the pressor response to infused norepinephrine. *Hypertension* 10:564–69
204. Anderson NB, Myers HF, Dickering T, Jackson JS. 1989. Hypertension in blacks: psychosocial and biological perspectives. *J. Hypertens.* 7:161–72
205. Calhoun DA. 1992. Hypertension in blacks: socioeconomic stress and sympathetic nervous system activity. *Am. J. Med. Sci.* 304:306–11
206. Sherwood A, Hinderliter AL. 1993. Responses to α - and β -adrenergic receptor agonists: effects of race in borderline hypertension compared to normotensive man. *Am. J. Hypertens.* 6:630–35
207. Stein CM, Lang CC, Singh I, He H, Wood AJJ. 1998. Increased vasoconstriction and decreased vasodilation not increased sympathetic activity are mechanisms for enhanced vascular reactivity in African-Americans. *Circulation* 98:I472 (Abstr.)
208. Stein CM, Lang CC, Singh I, Wood AJJ. 1998. Increased vascular alpha-adrenergic sensitivity in African-Americans. *Clin. Pharmacol. Ther.* 63:176 (Abstr.)
209. Eichler HG, Blaschke TF, Hoffman BB. 1990. Decreased responsiveness of superficial hand veins to phenylephrine in black normotensive males. *J. Cardiovasc. Pharmacol.* 16:177–81
210. de Mey J, Vanhoutte PM. 1981. Uneven distribution of postjunctional α_1 - and α_2 -like adrenoceptors in canine arterial and venous smooth muscle. *Circ. Res.* 48:875–84
211. Büscher R, Herrmann V, Insel PA. 1999. Human adrenoceptor polymorphisms: evolving recognition of clinical

- importance. *Trends Pharmacol. Sci.* 20: 94–99
212. Büscher R, Herrmann V, Ring KM, Kailasam MT, O'Lonnor DT, et al. 1999. Variability in phenylephrine response and essential hypertension: a search for human α_{1B} -adrenergic receptor polymorphisms. *J. Pharmacol. Exp. Ther.* 291:793–98
213. Xie HG, Kim RB, Stein CM, Gainer JV, Brown NJ, et al. 1999. α_{1A} -Adrenergic receptor polymorphism: association with ethnicity but not essential hypertension. *Pharmacogenetics* 9:651–56
214. Bylund DB, Eikenberg DC, Hieble JP, Langer SZ, Lefkowitz RJ, et al. 1994. IV. International Union of Pharmacology Nomenclature of Adrenoceptors. *Pharmacol. Rev.* 46:121–36
215. Hoffman BB, Lefkowitz RJ, Taylor P. 1996. Neurotransmission: The autonomic somatic and motor nervous system. In *Goodman and Gilman's the Pharmacological Basis of Therapeutics*, eds JG Hardman, LE Limbird, 9th ed, pp. 105–39. New York: McGraw-Hill
216. McDevitt DG. 1989. In vivo studies on the function of cardiac β -adrenoceptors in man. *Eur. Heart J.* 10(suppl B):22–28
217. Brodde O-E. 1991. β_1 - and β_2 -Adrenoceptors in the human heart: properties, function, and alterations in chronic heart failure. *Pharmacol. Rev.* 43:203–42
218. Zhou HH, Koshakji RP, Silberstein DJ, Wilkinson GR, Wood AJJ. 1989. Racial differences in drug response: altered sensitivity to and clearance of propranolol in men of Chinese descent as compared with American whites. *N. Engl. J. Med.* 320:560–70
219. Zhou HH, Wood AJJ. 1991. Increased suppression of exercise-induced increase in renin by propranolol in Chinese subjects. *Clin. Pharmacol. Ther.* 50:150–56
220. Johnson JA, Burlew BS, Stiles RN. 1995. Racial differences in β -adrenoceptor-mediated responsiveness. *J. Cardiovasc. Pharmacol.* 25:90–96
221. Richardson AD, Piepho RW. 2000. Effect of race on hypertension and antihypertensive therapy. *Intl. J. Clin. Pharmacol. Ther.* 38:75–79
222. Anonymous. Veterans Administration Cooperative Study Group on Antihypertensive Agents. 1982. Comparison of propranolol and hydrochlorothiazide for the initial treatment of hypertension. I. Results of short-term titration with emphasis on racial differences in response. *JAMA* 248:1996–2003
223. Anonymous. Veterans Administration Cooperative Study Group on Antihypertensive Agents. 1983. Efficacy of nadolol alone and combined with bendroflumethiazide and hydralazine for systemic hypertension. *Am. J. Cardiol.* 52:1230–37
224. Materson BJ, Reda DJ, Cushman WC, Massie BM, Freis ED, et al. 1993. Single-drug therapy for hypertension in men: a comparison of six antihypertensive agents with placebo. *N. Engl. J. Med.* 328:914–21
225. Seedat YK. 1989. Varying responses to hypotensive agents in different racial groups: black versus white differences. *J. Hypertens.* 7:515–18
226. Cushman WC, Reda DJ, Perry HM Jr, Williams D, Abdellatif M, et al. 2000. Regional and racial differences in response to antihypertensive medication use in a randomized controlled trial of men with hypertension in the United States. *Arch. Intern. Med.* 160:825–31
227. Frielle T, Collins S, Kiefer WD, Caron MG, Lefkowitz RJ, et al. 1987. Cloning of the cDNA from the human β_1 -adrenergic receptor. *Proc. Natl. Acad. Sci. USA* 84:7920–42
228. Tesson F, Charron P, Peuchmaurd M, Nicaud V, Cambien F, et al. 1999. Characterization of a unique genetic variant in the β_1 -adrenoceptor gene and evaluation

- of its role in idiopathic dilated cardiomyopathy. *J. Mol. Cell. Cardiol.* 31:1025–32
229. Podlowski S, Wenzel K, Luther HP, Müller J, Bramlage P, et al. 2000. β_1 -Adrenoceptor gene variations: a role in idiopathic dilated cardiomyopathy? *J. Mol. Med.* 78:87–93
230. Börjesson M, Magnusson Y, Andersson B. 1999. A novel polymorphism in the gene coding for the β_1 -receptor associated with survival in patients with heart failure. *J. Am. Coll. Cardiol.* 33:261A (Abstr.)
231. Maqbool A, Hall AS, Ball SG, Balmfort AJ. 1999. Common polymorphisms of β_1 -adrenoceptor: identification and rapid screening assay. *Lancet* 353:897
232. Mason DA, Moore JD, Green SA, Liggett SB. A gain-of-function polymorphism in a G-protein coupling domain of the human β_1 -adrenergic receptor. *J. Biol. Chem.* 274:12670–74
233. Moore JD, Mason DA, Green SA, Hsu J, Liggett SB. 1999. Racial differences in the frequencies of cardiac β_1 -adrenergic receptor polymorphisms: analysis of c145A→G and c1165G→C. *Hum. Mutat.* 14:271
234. Xie HG, Kim RB, Dishy V, Sofowora G, Xiao ZS, et al. 2000. Ethnic differences in beta-1 adrenoceptor polymorphism(Arg³⁸⁹→Gly) frequency in African-Americans, Caucasians and Chinese. *Clin. Pharmacol. Ther.* 67:125 (Abstr.)
- 234a. Xie HG, Dishy V, Sofowora G, Kim RB, Wood AJJ, et al. 2001. Human beta-1 adrenergic receptor polymorphism (Arg389Gly) is not associated with essential hypertension in black or white Americans. *Clin. Pharmacol. Ther.* In press (Abstr.)
- 234b. Xie HG, Dishy V, Sofowora G, Kim RB, Landau R, et al. 2001. Arg389Gly beta-1 adrenoceptor polymorphism varies in frequency among different ethnic groups but does not alter response in vivo. *Pharmacogenetics* In press
235. Kobilka BK, Dixon RAF, Frielle T, Dohlman HG, Bolanowski MA, et al., 1987. cDNA for the human β_2 -adrenergic receptor: a protein with multiple membrane-spanning domains and encoded by a gene whose location is shared with that of the receptor of platelet derived growth factor. *Proc. Natl. Acad. Sci. USA* 84:46–50
236. Kapoor C, Singarajah C, Zafar H, Adubofour KO, Takahashi B, et al. 1996. Impaired β_2 -adrenergic agonist-induced venodilation in Indians of Asian origin. *Clin. Pharmacol. Ther.* 59:569–76
237. Lang CC, Stein CM, Brown RM, Deegan R, Nelson R, et al. 1995. Attenuation of isoproterenol-mediated vasodilation in blacks. *N. Engl. J. Med.* 333:155–60
238. Watkins LL, Dimsdale JE, Ziegler MG. 1995. Reduced β_2 -receptor mediated vasodilation in African Americans. *Life Sci.* 57:1411–16
239. Stein CM, Lang CC, Nelson R, Brown M, Wood AJJ. 1997. Vasodilation in black Americans: attenuated nitric oxide-mediated responses. *Clin. Pharmacol. Ther.* 62:436–43
240. Cardillo C, Kilcoyne CM, Cannon III RO, Panza JA. 1999. Attenuation of cyclic nucleotide-mediated smooth muscle relaxation in blacks as a cause of racial differences in vasodilator function. *Circulation* 99:90–95
241. Dawes M, Chowieńczyk PJ, Ritter JM. 1997. Effects of inhibition of the L-arginine/ nitric oxide pathway on vasodilation caused by β -adrenergic agonists in human forearm. *Circulation* 95:2293–97
242. Lembo G, Iaccarino G, Vecchione C, Barbato E, Izzo R, et al., 1997. Insulin modulation of an endothelial nitric oxide component present in the α_2 - and β -adrenergic responses in human forearm. *J. Clin. Invest.* 100:2007–14
243. Vecchione C, Izzo R, Fontana D, Fratta L,

- de Santis D, et al. 1998. Nitric oxide component present in β -adrenergic vasodilation is impaired in essential hypertension. *Am. J. Hypertens.* 11:164A (Abstr.)
244. Cardillo C, Kilcoyne CM, Quyyumi AA, Cannon RO III, Panza JA. 1997. Decreased vasodilator response to isoproterenol during nitric oxide inhibition in humans. *Hypertension* 30:918–21
245. Liggett S. 1995. Functional properties of human β_2 -adrenergic receptor polymorphisms. *News Physiol. Sci.* 10:265–73
246. Green SA, Turki J, Innis M, Liggett SB. 1994. Amino-terminal polymorphisms of the human β_2 -adrenergic receptor impart distinct agonist-promoted regulatory properties. *Biochemistry* 33:9414–19
247. Green SA, Turki J, Bejarano P, Hall IP, Liggett SB. 1995. Influence of β_2 -adrenergic receptor genotypes on signal transduction in human airway smooth muscle cells. *Am. J. Respir. Cell Mol. Biol.* 13:25–33
248. Chong LK, Chowdry J, Ghahramani P, Peachell PT. 2000. Influence of genetic polymorphisms in the β_2 -adrenoceptor on desensitization in human lung mast cells. *Pharmacogenetics* 10:153–62
- 248a. Dishy V, Sofowora GG, Xie HG, Stein CM, Kim RB, et al. 2000. Agonist-mediated vasodilation and desensitization in vivo is determined by β_2 -adrenergic receptor polymorphisms. *Circulation* 102(18):102 (Abstr.)
249. Gratze G, Fortin J, Labugger R, Binder A, Kotanko P, et al. 1999. β -2 Adrenergic receptor variants affect resting blood pressure and agonist-induced vasodilation in young adult Caucasians. *Hypertension* 33:1425–30
250. Hoit BD, Suresh DP, Craft L, Walsh RA, Liggett SB. 2000. β_2 -Adrenergic receptor polymorphisms at amino acid 16 differentially influence agonist-stimulated blood pressure and peripheral blood flow in normal individuals. *Am. Heart J.* 139:537–42
251. Hall IP, Gazis AG, White DJ, Wheatley AP, Cockcroft JR. 1998. Association of β_2 -adrenoceptor polymorphism with vascular reactivity in humans. *Br. J. Clin. Pharmacol.* 45:510P (Abstr.)
252. Xie HG, Stein CM, Kim RB, Xiao ZS, He N, et al. 1999. Frequency of functionally important beta-2 adrenoceptor polymorphisms varies markedly among African-American, Caucasian and Chinese individuals. *Pharmacogenetics* 9:511–16
253. Xie HG, Stein CM, Kim RB, Gainer JV, Sofowora G, et al., 2000. Human β_2 -adrenergic receptor polymorphisms: no association with essential hypertension in black or white Americans. *Clin. Pharmacol. Ther.* 67:670–75
254. Moncada S, Higgs A. 1993. The L-arginine-nitric oxide pathway. *N. Engl. J. Med.* 329:2002–12
255. Vallance P, Collier J, Moncada S. 1989. Effects of endothelium-derived nitric oxide on peripheral arteriolar tone in man. *Lancet* 2:997–1000
256. Vallance P. 1998. Nitric oxide in the human cardiovascular system-SKB lecture 1997. *Br. J. Clin. Pharmacol.* 45:433–39
257. Cardillo C, Kilcoyne CM, Cannon III RO, Panza JA. 1998. Racial differences in nitric oxide-mediated vasodilator response to mental stress in the forearm circulation. *Hypertension* 31:1235–39
258. Jones DS, Andrawis NS, Abernethy DR. 1999. Impaired endothelial-dependent forearm vascular relaxation in black Americans. *Clin. Pharmacol. Ther.* 65: 408–12
259. Gainer JV, Stein M, King D, Brown NJ. 1998. Racial differences in bradykinin-induced forearm blood flow. *Circulation* 98:1243 (Abstr.)
260. Marsden PA, Heng HHQ, Scherer SW, Stewart RJ, Hall AV, et al. 1993. Structure and chromosomal localization of the human constitutive endothelial nitric oxide

- synthase gene. *J. Biol. Chem.* 268:17478–88
261. Babaoglu MO, Abernethy DR. 2000. Polymorphic variant of endothelial nitric oxide synthase (eNOS) markedly impairs endothelium-dependent vascular relaxation. *Clin. Pharmacol. Ther.* 67:141 (Abstr.)
262. Philip I, Plantefevre G, Vuillaumier-Barrot S, Vicaut E, LeMarie C, et al. 1999. G894T polymorphism in the endothelial nitric oxide synthase gene is associated with an enhanced vascular responsiveness to phenylephrine. *Circulation* 99:3096–98
263. Xie HG, Stein CM, Kim RB, Sofowora G, Dishy V, et al. 2000. Frequency of endothelial nitric oxide synthase Glu²⁹⁸Asp polymorphism varies among African-Americans, Caucasians, and Chinese. *Clin. Pharmacol. Ther.* 67:125 (Abstr.)
264. Siffert W, Rosskopf D, Siffert G, Busch S, Moritz A, et al. 1998. Association of a human G-protein $\beta 3$ subunit variant with hypertension. *Nature Genet.* 18:45–48
265. Ansari-Lari MA, Muzny DM, Lu J, Lu F, Lilley CE, et al. 1996. A gene-rich cluster between the CD4 and triosephosphate isomerase genes at human chromosome 12p13. *Genome Res.* 6:314–26
266. Schunkert H, Hense H-W, Döring A, Riegger GAJ, Siffert W. 1998. Association between a polymorphism in the G protein $\beta 3$ subunit gene and lower renin and elevated diastolic blood pressure levels. *Hypertension* 32:510–13
267. Beige J, Hohenbleicher H, Distler A, Sharma AM. 1999. G-protein $\beta 3$ subunit C825T variant and ambulatory blood pressure in essential hypertension. *Hypertension* 33:1049–51
268. Benjafield AV, Jeyasingam CL, Nyholt DR, Griffiths LR, Morris BJ. 1998. G-protein $\beta 3$ subunit gene (*GNB3*) variant in causation of essential hypertension. *Hypertension* 32:1094–7
269. Dong Y, Zhu H, Sagnella GA, Carter ND, Cook DG, et al. 1999. Association between the C825T polymorphism of the G protein $\beta 3$ -subunit gene and hypertension in blacks. *Hypertension* 34:1193–6
270. Kato N, Sugiyama T, Morita H, Kurihara H, Yamori Y, et al. 1998. G protein $\beta 3$ subunit variant and essential hypertension in Japanese. *Hypertension* 32:935–38
271. Kario K, Fujiwara M, Sone Y, Saiki K, Hoshida S, et al. 1999. G protein $\beta 3$ subunit gene variant, twenty-four-hour blood pressure, and hypertensive cerebrovascular disease in a Japanese population. *Am. J. Hypertens.* 12:1159–60
272. Ishikawa K, Imai Y, Katsuya T, Ohkubo T, Tsuji I, et al. 2000. Human G-protein $\beta 3$ subunit variant is associated with serum potassium and total cholesterol levels but not with blood pressure. *Am. J. Hypertens.* 13:140–45
273. Hegele RA, Harris SB, Hankey AJG, Cao H, Zinman B. 1998. G protein $\beta 3$ subunit gene variant and blood pressure variation in Canadian Oji-Cree. *Hypertension* 32:688–92
274. Brand E, Herrmann S-M, Nicaud V, Ruidavets J-B, Evans A, et al. 1999. The 825C/T polymorphism of the G-protein subunit $\beta 3$ is not related to hypertension. *Hypertension* 33:1175–8
275. Larson N, Hutchinson R, Boerwinkle E. 2000. Lack of association of 3 functional gene variants with hypertension in African Americans. *Hypertension* 35:1297–1300
276. Poch E, González D, Gómez-Angelats E, Enjuto M, Paré JC, et al. 2000. G-protein $\beta 3$ subunit gene variant and left ventricular hypertrophy in essential hypertension. *Hypertension* 35:214–18
277. Jacobi J, Hilgers KF, Schlaich MP, Siffert W, Schmieder RE. 1999. 825T allele of the G-protein $\beta 3$ subunit gene (*GNB3*) is associated with impaired left ventricular diastolic filling in essential hypertension. *J. Hypertens.* 17:1457–62

278. Siffert W, Forster P, Jöckel K-H, Mvere DA, Brinkmann B, et al. 1999. World-wide ethnic distribution of the G protein $\beta 3$ subunit 825T allele and its association with obesity in Caucasian, Chinese, and black African individuals. *J. Am. Soc. Nephrol.* 10:1921–30
279. Siffert W, Naber C, Walla M, Ritz E. 1999. G protein $\beta 3$ subunit 825T allele and its potential association with obesity in hypertensive individuals. *J. Hypertens.* 17:1095–98
280. Hegele RA, Anderson C, Young TK, Connelly PW. 1999. G-protein $\beta 3$ subunit gene splice variant and body fat distribution in Nunavut Inuit. *Genome Res.* 9:972–77
281. Hoche B, Slowinski T, Stolze T, Pleschka A, Neumayer H-H, et al. 2000. Association of maternal G protein $\beta 3$ subunit 825T allele with low birth-weight. *Lancet* 355:1241–42
282. Gutersohn A, Naber C, Müller N, Erbel R, Siffert W. 2000. G protein $\beta 3$ subunit 825 TT genotype and post-pregnancy weight retention. *Lancet* 355:1240–41
- 282a. Gupta S, Dishy V, Xie HG, Landau R, Smiley RM, et al. 2001. G-protein $\beta 3$ -subunit 825C/T polymorphism is associated with weight gain in pregnancy but not pre-eclampsia. *Clin. Pharmacol. Ther.* In press (Abstr.)
283. Limbird LE. 1988. Receptors linked to the inhibition of adenylate cyclase: additional signaling mechanisms. *FASEB J.* 2:1762–67
284. Nebigil C, Malik KU. 1992. α -Adrenergic receptor subtypes involved in prostaglandin synthesis are coupled to Ca^{2+} channels through pertussis toxin-sensitive guanine nucleotide-binding protein. *J. Pharmacol. Exp. Ther.* 266:1113–24
285. Baumgart D, Naber C, Haude M, Oldenburg O, Erbel R, et al. 1999. G protein $\beta 3$ subunit 825T allele and enhanced coronary vasoconstriction on α_2 -adrenoceptor activation. *Circ. Res.* 85:965–69.
286. Schäfers RF, Nürnberger J, Rütz A, Siffert W, Philipp TH, et al. 1999. Vasoconstrictor response of the human dorsal hand vein in young normotensive men carrying the 825T-allele of the G-protein $\beta 3$ subunit. *Br. J. Clin. Pharmacol.* 48:872P (Abstr.)
287. Weinshilboum RM, Otterness DM, Szumlanski CL. 1999. Methylation pharmacogenetics: catechol *O*-methyltransferase, thiopurine methyltransferase, and histamine *N*-methyltransferase. *Annu. Rev. Pharmacol. Toxicol.* 39:19–52
288. Nagata K, Yamazoe Y. 2000. Pharmacogenetics of sulfotransferase. *Annu. Rev. Pharmacol. Toxicol.* 40:159–76
289. Tukey RH, Strassburg CP. 2000. Human UDP-glucuronosyltransferases: metabolism, expression, and disease. *Annu. Rev. Pharmacol. Toxicol.* 40:581–616
290. Hein DW, Doll MA, Fretland AJ, Leff MA, Webb SJ, et al. 2000. Molecular genetics and epidemiology of the NAT1 and NAT2 acetylation polymorphism. *Cancer Epidemiol. Biomark. Prev.* 9:29–42